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THE PHARMACOKINETICS OF METHSUXIMIDE  
AND A MAJOR METABOLITE IN THE DOG

BY

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THE PHARMACOKINETICS OF METHSUXIMIDE  
AND A MAJOR METABOLITE IN THE DOG

Michael R. Dobrinska

(Under the supervision of Associate Professor Peter G. Welling)

Methsuximide is an agent useful in the treatment of petit mal epilepsy and is marketed as Celontin<sup>R</sup> 150 or 300 mg oral capsules. One of its major biotransformation products identified in blood is the N-demethylated compound, 2-methyl-2-phenylsuccinimide. This metabolite is pharmacologically active and potentially toxic. Methsuximide induces its own metabolism by stimulating the activity of drug metabolizing enzymes. Little information exists on the pharmacokinetic behavior of this drug in humans or experimental animals. The present studies were undertaken to elucidate the pharmacokinetics of methsuximide after single intravenous doses.

The dog was used as an experimental model since intravenous doses of methsuximide or metabolite were required to best define distribution volumes. It was found that methsuximide obeys the two compartment open model while the metabolite, after intravenous dosing, was best described by the one compartment open model. The metabolite plasma level profile after methsuximide dosing was described by a triexponential equation showing the influence of the methsuximide biexponential and its own monoexponential equations. The blood level data were fit to these equations with the aid of

a digital computer. Parameters reflecting the distribution and elimination characteristics of parent drug and metabolite were obtained. The metabolite's biological half-life was found to exceed that of the parent drug by a factor of about 15. The N-demethylation metabolic step accounted for approximately 40% of the overall elimination of methsuximide.

APPROVED

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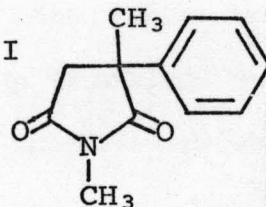
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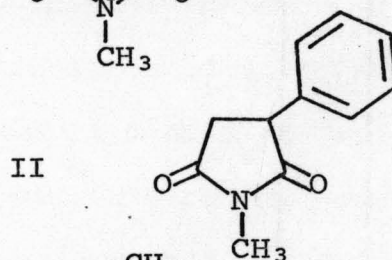
## INTRODUCTION

Succinimide derivatives potentially useful in the treatment of petit mal epilepsy were first introduced by Miller and Long in 1951 (1). The anticonvulsant activity of the  $\alpha$ -phenylsuccinimides was confirmed by Chen *et al.* in 1951 (2). It was found that the  $\alpha$ -phenyl, 2-methyl and N-methyl groups on the succinimide ring increased anticonvulsant activity while an increase in length of the N-alkyl side chain decreased activity. Another derivative, devoid of the  $\alpha$ -phenyl group, proved also to be highly efficacious in preliminary clinical studies (3, 4). Those succinimides currently in use in the United States are:

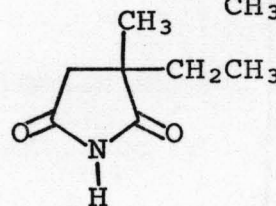
(+)-N,2-dimethyl-2-phenylsuccinimide I  
(méthsuximide, Celontin<sup>®</sup>)<sup>a</sup>



(+)-N-methyl-2-phenylsuccinimide II  
(phensuximide, Milontin<sup>®</sup>)



(+)-2-ethyl-2-methylsuccinimide III  
(ethosuximide, Zarontin<sup>®</sup>)



There is little quantitative information on the pharmacokinetics of the succinimides in experimental animals or

<sup>a</sup>Celontin, Milontin and Zarontin are registered trademarks of Parke-Davis and Company. In all drug studies referred to, or studied in this thesis, the racemic modification was used.

man. Methsuximide possesses many interesting properties which affect its pharmacokinetic profile and was chosen as the drug used in the present studies.

Glazko and Dill (5) have reported methsuximide plasma levels in twelve normal adult males at two dose levels of 0.6 g and 1.2 g in a cross-over design study. Mean peak plasma levels of  $3.1 \pm 0.5$  and  $6.8 \pm 0.6$  (standard error)  $\mu\text{g ml}^{-1}$  occurred at one hour after dosing 0.6 and 1.2 g respectively. Thus, in this dosage range the plasma level was found to be roughly proportional to dose. The mean plasma half-life,  $t_{1/2}$ , with both doses was 2.6 hours over the 1-12 hour period. The experiments were repeated weekly for four weeks. It was found that the plasma  $t_{1/2}$ 's during the first week were approximately 75% greater than during the fourth week. This was interpreted in terms of induction of hepatic drug metabolizing enzymes. Recent evidence confirmed this interpretation since administration of I, II or III for three days to the rat resulted in increased oxidation of hexobarbitone, hydroxylation of aniline, liver:body weight ratio, hepatic microsomal cytochrome P-450, hepatic  $\delta$ -amino-laevulinic acid synthetase and a reduction in hexobarbitone-induced hypnosis (6). These investigators also noted a decreased anticonvulsant activity after three days of methsuximide administration to the rat.

In 1971 Nicholls and Orton (7) reported on the absorption, distribution, metabolism and excretion (ADME) of  $\text{N-}^{14}\text{CH}_3$  labeled methsuximide in the rat. After oral dosing

they found significant levels of radioactivity in the brain, liver, kidney, heart, adrenals, spleen, testes, lungs, salivary glands, eyes, fat and skeletal muscle indicating rapid and extensive drug distribution. Levels in body fat tended to be higher than those in other tissues. About 29% of the administered dose appeared as  $^{14}\text{CO}_2$  in expired air within 24 hours after dosing. This suggested that there may be extensive N-demethylation of the parent drug to 2-methyl-2-phenylsuccinimide. Rats which received oral  $100 \text{ mg kg}^{-1}$  methsuximide doses exhibited anti-leptazol activity for up to six hours after dosing yet there was little parent drug in the brain at this time. It was suggested that since the metabolite also possesses anti-leptazol activity (2, 7, 8) it may have been responsible for the extended pharmacological activity. This is not uncommon among antiepileptic drugs as a similar phenomenon exists after mephobarbital, metharbital, mesantoin, trimethadione or paramethadione administration (9).

Few studies on the metabolism of methsuximide have appeared in the literature. Horning and coworkers (10) found N,2-dimethyl-2-(3,4-dihydroxy-1,5-cyclohexadien-1-yl)-succinimide, N,2-dimethyl-2-(4-hydroxyphenyl)-succinimide, N,2-dimethyl-2-(3-hydroxyphenyl)-succinimide, N-methyl-2-hydroxymethyl-2-phenylsuccinimide, two isomeric N,2-dimethyl-3-hydroxy-2-phenylsuccinimides and a diol presumed to be N,2-dimethyl-2-(4-hydroxyphenyl)-3-hydroxysuccinimide as well as small amounts of parent drug in the urine of the rat, guinea pig and man after oral methsuximide administration.

Assignment of metabolite structures was based on gas chromatography-mass spectrometry (GC-MS) after methylation and trimethylsilylation without pure metabolite reference standards. Dudley *et al.* (11) studied the metabolism of methsuximide in the dog and identified metabolites in 48 hour urine by melting point determination, mass spectroscopy, IR and UV absorption spectroscopy and NMR using pure synthesized metabolites as reference standards. After dosing one or two grams of methsuximide, roughly 7-20% of the dose was recovered as 2-(p-hydroxyphenyl)-2-methylsuccinimide and N-methyl-2-(p-hydroxyphenyl)-2-methylsuccinimide. The former derivative was the primary para-hydroxylated metabolite. Only a small amount of the N-demethylated metabolite was recovered as was the case after 2-methyl-2-phenylsuccinimide itself was administered. There was no evidence of metabolites formed by opening of the succinimide ring. No unchanged methsuximide was recovered and the authors stated that the N-demethylation reaction is probably the primary biotransformation route for methsuximide.

Nicholls and Orton (8) discussed methsuximide ADME in quantitative terms. The absorption half-life after oral doses in the rat was 17.4 minutes while distribution characteristics were similar to those previously found (7). The 2-methyl-2-phenylsuccinimide metabolite was identified in the urine of the rat, guinea pig (12) and man after oral methsuximide administration. Although the anticonvulsant activity of the N-demethyl metabolite is about half that of

the parent drug it has been found to be active for up to six hours after oral metabolite administration to the rat (2). The metabolite has been identified in rabbit plasma by GC-MS after oral methsuximide dosing (13). The parent drug was rapidly metabolized and within 30 minutes the concentration of the metabolite was much higher than that of the drug.

The plasma of 17 patients on chronic methsuximide therapy was analyzed for methsuximide and 2-methyl-2-phenylsuccinimide using the sensitive quadrupole mass fragmentography method (14). The metabolite level was found to be 700 times that of the parent drug. Metabolite levels of less than  $10 \mu\text{g ml}^{-1}$  were ineffective in seizure control while levels in excess of  $40 \mu\text{g ml}^{-1}$  were toxic. This metabolite is very lipophilic and has a  $\text{pK}_a$  of 8.52 (8). Since only 2.7% of the methsuximide dose was excreted into the 24 hour rat urine as the metabolite, while 29% was exhaled as  $^{14}\text{CO}_2$  in the same time period (8), a long metabolite biological  $t_{1/2}$  may be expected. Hence, the metabolite is not only rapidly formed, pharmacologically active and potentially toxic, but also may be persistent in the body. The clinical implications of this situation are obvious.

In 1973 Karch reported an overdose of methsuximide by an 18 year old female patient who ingested about 10 g of the drug (15). Sixteen hours post-ingestion plasma levels of drug and metabolite were 18 and  $44 \mu\text{g ml}^{-1}$ , respectively. At 64 hours post-ingestion methsuximide levels had declined to  $2 \mu\text{g ml}^{-1}$  whereas metabolite levels were only slightly

lowered at  $38 \mu\text{g ml}^{-1}$ . The clinical course was biphasic and consisted of initial lethargy, from which the patient could be easily aroused, a period of improvement followed by a long period of profound coma. Since the  $t_{1/2}$  of methsuximide is less than 3 hours the coma could not have been due to parent drug and was attributed to the 2-methyl-2-phenylsuccinimide metabolite. The biphasic toxicity experienced with this drug is potentially dangerous and requires close scrutiny in the treatment of overdose victims. This case report emphasizes the need for pharmacokinetic information not only on the parent drug but also on the metabolite. This information would also be invaluable in seeking solutions to problems associated with a drug which exhibits auto-induction of hepatic metabolizing enzymes and forms active and potentially toxic metabolites.

The present studies were designed to elucidate the pharmacokinetics of both methsuximide and 2-methyl-2-phenylsuccinimide after intravenous administration of these compounds to male beagle dogs.

Intravenous dosing was employed in order to best define the distribution and elimination characteristics of these agents in the absence of complications due to absorption rates or variability in bioavailability.

The use of beagle dogs may be rationalized in terms of the following: (1) dogs are often useful models in predicting the pharmacokinetic characteristics of a drug in humans (16); (2) intravenous methsuximide and

2-methyl-2-phenylsuccinimide are not officially recognized dosage forms for human use; (3) multiple blood samples over prolonged time periods were required to adequately define the pharmacokinetic profiles.

It is hoped that this thesis will provide some of the basic information necessary to improve methsuximide drug therapy in patients suffering from petit mal epilepsy.

## EXPERIMENTAL

A. Materials

Pure 5 gram samples of methsuximide and phensuximide were obtained from the manufacturer<sup>a</sup> while 3 grams of pure 2-methyl-2-phenylsuccinimide were obtained commercially<sup>b</sup>. Two male beagle dogs<sup>c</sup> were used; dog A weighed 14 kg and dog B 17.5 kg. The dogs had free access to water at all times but food was withheld a minimum of 12 hours prior to an experiment.

B. Experimental Procedure

The dog was prepared for each experiment by placing it in a restraining apparatus which consisted of a strong sheet which supported its body and through which its limbs protruded. The limbs were held secure by pediatric limb restraints tied to the shell of the restraining apparatus as in Figure 1. The dog was made as comfortable as possible and was held in restraints only during the initial period of the experiment, usually 1-2 hours, when frequent blood samples were obtained.

Two vein infusion sets<sup>d</sup> were positioned, one in a front

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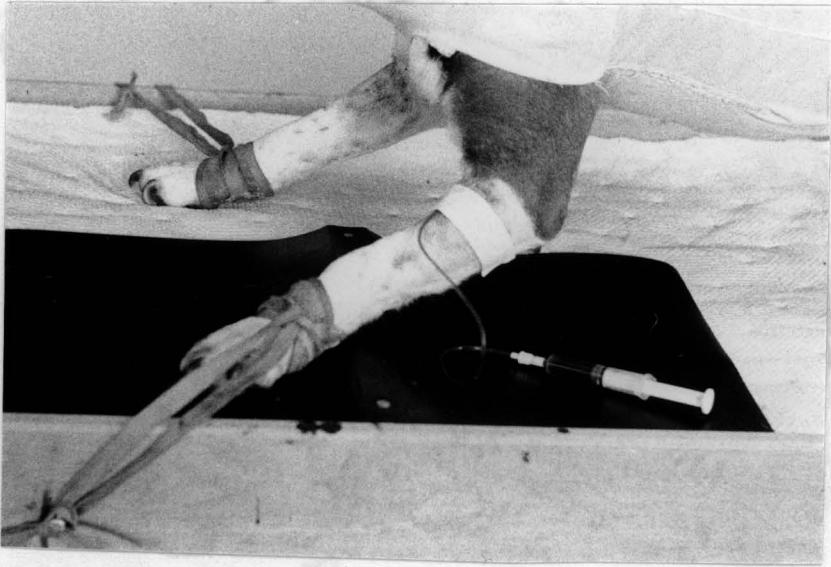
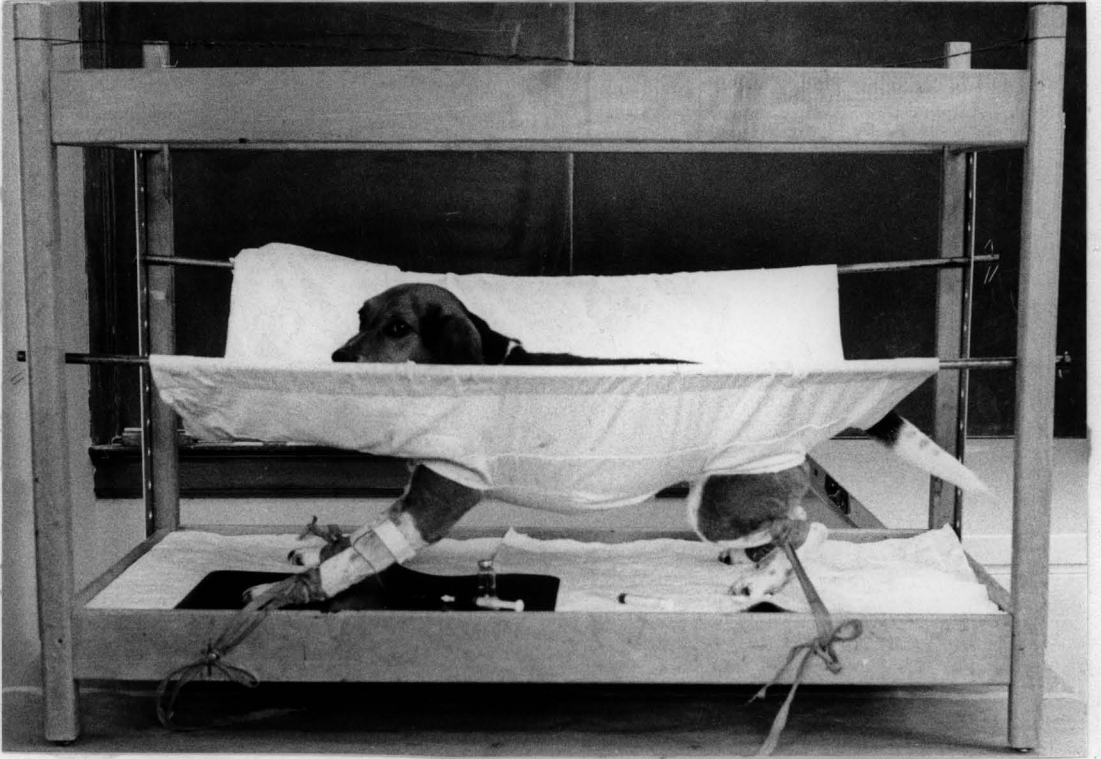
<sup>a</sup>Parke-Davis and Company, Detroit, Michigan.

<sup>b</sup>Custom synthesis, Aldrich Chemical Company, Milwaukee, Wisconsin.

<sup>c</sup>Animal Care Unit, the University of Wisconsin, Madison, Wisconsin.

<sup>d</sup>Miniset<sup>R</sup> Vein Infusion Set, Travenol Labs, Deerfield, Illinois.

Figure 1. Dog restraining apparatus (upper) showing limb restraints and vein infusion set placement (lower).



Photographs by Shabbir T. Anik.

limb vein, the other in the other front or a hind limb vein. The sets had 19 gauge X 7/8" needles with 30 cm of flexible plastic tubing attached which could contain about 0.6 ml of fluid. The infusion sets were kept free of blood clots by infusion of about 2 ml of sodium heparin solution<sup>a</sup> (10 units ml<sup>-1</sup>) into them approximately every hour they were in position. One set was used to rapidly administer the drug solution while the other was used to collect blood samples. Infusion sets were found to be necessary to assure intravenous dosing and to facilitate rapid collection of several blood samples in the first hour of the experiments without multiple venipunctures.

The water solubility of methsuximide is 2.8 mg ml<sup>-1</sup> at pH 7.0 and 25° C (5). A dose of 250 mg of either methsuximide or the metabolite was selected since this represents a low therapeutic dose in children. One assumption in the pharmacokinetic analysis was that the injection was accomplished instantaneously. The volume of drug solution which could be injected had therefore to be minimized and this precluded the use of an aqueous drug solution. The following co-solvent system was developed which yielded a visually clear solution of methsuximide or metabolite at 25° C:

methsuximide or,	
2-methyl-2-phenylsuccinimide	300 mg
propylene glycol, USP	4.5 ml
water for injection, USP qs ad.	6.0 ml

<sup>a</sup>Saline diluted Lipo-Heparin<sup>R</sup> 5000 u ml<sup>-1</sup>, Riker Labs, Northridge, California.

Injection of 5.0 ml of this solution provided 250 mg of drug. The drug solution was prepared within 2 hours of use and stored in a 10 ml sterile multi-dose vial. Before injecting, the solution was warmed to about 37° C to prevent chills and consequent local vasoconstriction (17). The 5 ml injection was accomplished in approximately 120 seconds. A faster injection led to drug precipitation and formation of a local hematoma. The infusion set was immediately flushed with 10 ml of normal saline solution. The set was removed and analysis of water flushings indicated no methsuximide or metabolite remained in the set.

Blood samples (5 ml) were then collected via the other infusion set at 0, 2-5, 10, 15, 30, 45, 60 and 120 minutes. Collection of blood at 3, 4, 5, 6, 8, 10, 12, 18, 24, 36 and 48 hours was done by direct venipuncture in most cases. Alterations in times occurred depending on the experiment. Before each blood sample was drawn 5 ml of "residual fluid" was withdrawn to avoid the 0.6 ml of heparin solution contained in the set from diluting the blood sample. The 5 ml blood sample was then drawn and replaced by a 5 ml injection of normal saline solution. The residual fluid was re-injected followed by 1-2 ml of heparin solution 5 units ml<sup>-1</sup>. Direct venipunctures were done on any leg. Blood samples were placed in 10 ml heparinized vacutainers<sup>a</sup> and

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<sup>a</sup>Green stopper Vacutainer<sup>®</sup> evacuated glass tubes coated with 143 units sodium heparin, Becton-Dickinson, Rutherford, New Jersey.

centrifuged<sup>a</sup>. The plasma, about 2 ml, was deep frozen until analyzed, usually within 2 weeks. Some experiments required 90-100 ml of blood; however, visual inspection of the centrifuged blood indicated no decrease in the hematocrit during the sampling period.

The minimum time between experiments on the same dog was one month. This reduced the danger of changes in pharmacokinetic parameters due to prior drug exposure, i.e. enzyme induction or residual drug related materials in the body, and allowed for complete healing of the venipuncture sites.

### C. Assay Procedure

The assay was a modification of that of Kinkel et al. (5). One ml of plasma or 1 ml of standard aqueous methsuximide and metabolite standard in blank plasma was pipetted into 13 ml glass stoppered centrifuge tubes and acidified with 0.5 ml of 0.5 N HCl. Five mls of  $\text{CHCl}_3$ <sup>b</sup> was added and the tubes were horizontally shaken on a flat-bed shaker for 15 minutes. The tubes were centrifuged at 2300 rpm for 15 minutes or until clean phase separation was accomplished. Since emulsification was a frequent problem 4.0 ml aliquots of the clear  $\text{CHCl}_3$  drug containing layer were pipetted off and placed in dry clean tubes. One ml of phensuximide  $\text{CHCl}_3$  solution,  $6 \mu\text{g ml}^{-1}$ , was added to the  $\text{CHCl}_3$  aliquot as an

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<sup>a</sup>2300 rpm's for 15 minutes with the Dynac centrifuge, Clay Adams, Parsippany, New Jersey.

<sup>b</sup> $\text{CHCl}_3$ , A.R., Mallinckrodt, as supplied.

external standard. The  $\text{CHCl}_3$  was carefully evaporated to dryness in a  $40^\circ\text{C}$  water bath under a gentle stream of  $\text{N}_2$ . The residue was immediately redissolved in about 100  $\mu\text{l}$  of  $\text{CS}_2^{\text{a}}$  and the samples were analyzed on the same day by GC.

The GC<sup>b</sup> used for quantification was a programmable dual column gas chromatograph equipped with dual flame ionization detectors (FID). A six foot two mm (ID) U shaped glass column packed with 3% OV-17 on Gas Chrome Q 100/120 mesh<sup>c</sup> was used. The carrier gas was  $\text{N}_2$  flowing at 20  $\text{ml min}^{-1}$  while the flame gases were  $\text{H}_2$  and compressed air flowing at 20 and 200  $\text{ml min}^{-1}$ , respectively. The solid state electrometer was set at a range of  $10^{-11}$  amperes and the attenuator at 8-64 so that a recorder<sup>d</sup> full scale pen deflection occurred with an input signal of  $8-64 \times 10^{-11}$  amperes. The GC was operated in the single column mode at an isothermal column temperature of  $175^\circ\text{C}$  with FID bath at  $200^\circ\text{C}$  and injection block at  $210^\circ\text{C}$ . Sample volume introduced by the on column injection technique was 3  $\mu\text{l}$  and chart speed was 15 inches  $\text{hr}^{-1}$ . The column was initially conditioned for 48 hours at  $200^\circ\text{C}$  and overnight after each series of plasma sample extracts were run. In order to prevent compound adsorption at active sites and to maintain column integrity

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<sup>a</sup>Carbon disulfide, A.R., Mallinckrodt, as supplied.

<sup>b</sup>Nuclear Chicago Selectra System 5000.

<sup>c</sup>Applied Science Laboratories, State College, Pennsylvania.

<sup>d</sup>Barber-Colman Chromocorder Dual Pen X-Y Recorder, Rockford, Illinois.

20  $\mu$ l of Silyl-8<sup>a</sup> was injected onto the column after approximately every six hours of column use.

D. Summary of Experiments

The experiments conducted may be summarized as follows:

<u>Experiment</u>	<u>Dog</u>	<u>Dose</u>	
A	A	Methsuximide	250 mg
B	B	Methsuximide	250 mg
C	B	Methsuximide	250 mg
D	B	2-Methyl-2-phenylsuccinimide	250 mg
E	B	2-Methyl-2-phenylsuccinimide	250 mg

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<sup>a</sup>Pierce Chemical Company, Rockford, Illinois.

## RESULTS

A. Assay1. Plasma Sample Workup

The plasma sample workup was repeatedly changed in an effort to gain maximum extraction efficiency and cleaner samples for GC analysis. Drug recovery trials were done when  $\text{CHCl}_3$  was the final GC solvent. Triplicate runs of blank dog plasma containing methsuximide and metabolite at concentrations of 0.5, 1.0, 2.0, 4.0, 6.0, 8.0, 10.0, 15.0, 20.0, 30.0, 35.0 and 40.0  $\mu\text{g ml}^{-1}$ , representing the range in expected plasma levels, were done. The recovery of methsuximide was  $92 \pm 3\%$  while that of 2-methyl-2-phenylsuccinimide was  $88 \pm 5$  (S.D.)%. It has been reported (5) that evaporation of the  $\text{CHCl}_3$  layer to dryness may result in drug losses. However, improved quantitation of drug and metabolite peaks resulted from changing the final GC solvent from  $\text{CHCl}_3$  to  $\text{CS}_2$  which gives a negligible solvent peak. This change resulted in no significant differences in recoveries of either compound. All aqueous or  $\text{CHCl}_3$  reference solutions used in the recovery trials were refrigerated and found to remain stable for periods in excess of 3 months. Derivatization of drug samples was found not to be necessary for the GC analysis.

2. Gas Chromatography

Gas chromatography can be extremely sensitive and

reproducible if the operating parameters are carefully selected (19). The H<sub>2</sub>/air flame gas flow ratio should be 1:10 to obtain a good flame but the individual flows can be altered to obtain maximum sensitivity of the FID to the compounds. Figure 2 is a plot of area under the drug peaks, determined by disc integration, versus H<sub>2</sub> flow rate. The selection of flame gas flow rates of 20 and 200 ml min<sup>-1</sup>, yielded the maximum sensitivity of the FID to both compounds.

The optimum carrier gas (N<sub>2</sub>) flow rate was determined through use of the experimentally obtained Van Deemter plot (18, 19). A GC column may be considered as a series of "theoretical plates" which are of a thickness such that equilibrium is reached between solution leaving the plate and the average concentration of solute in the stationary phase (OV-17) throughout the plate (18). The thickness of the plate is referred to as the height equivalent of a theoretical plate (HETP) and is obtained through equations 1-3,

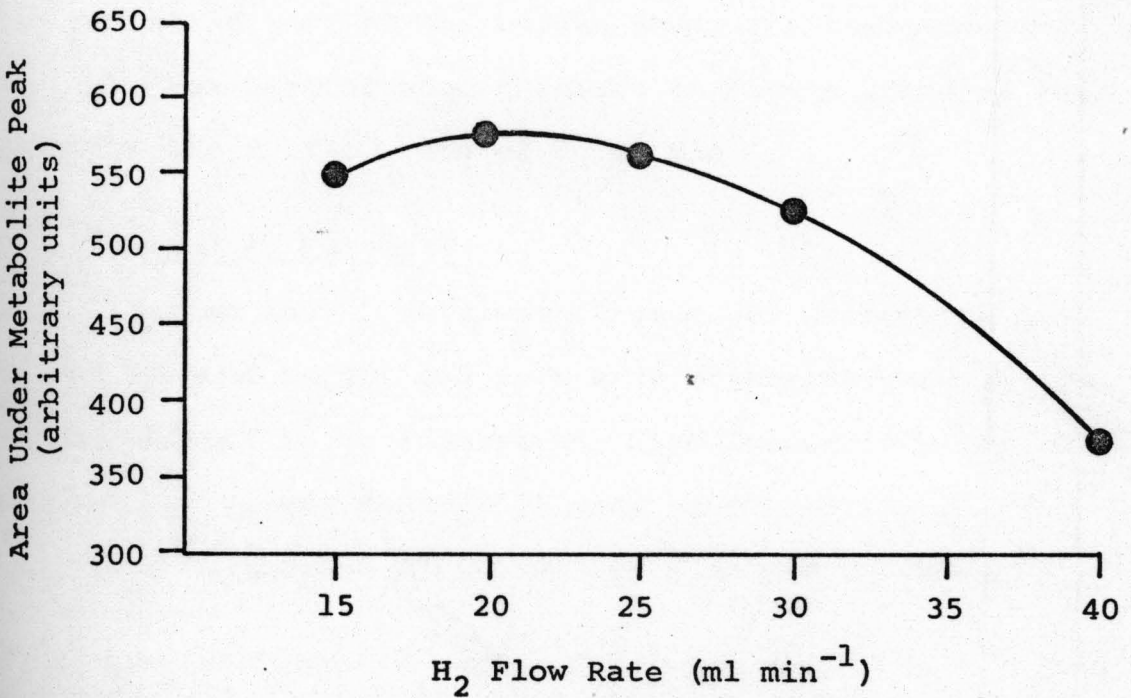
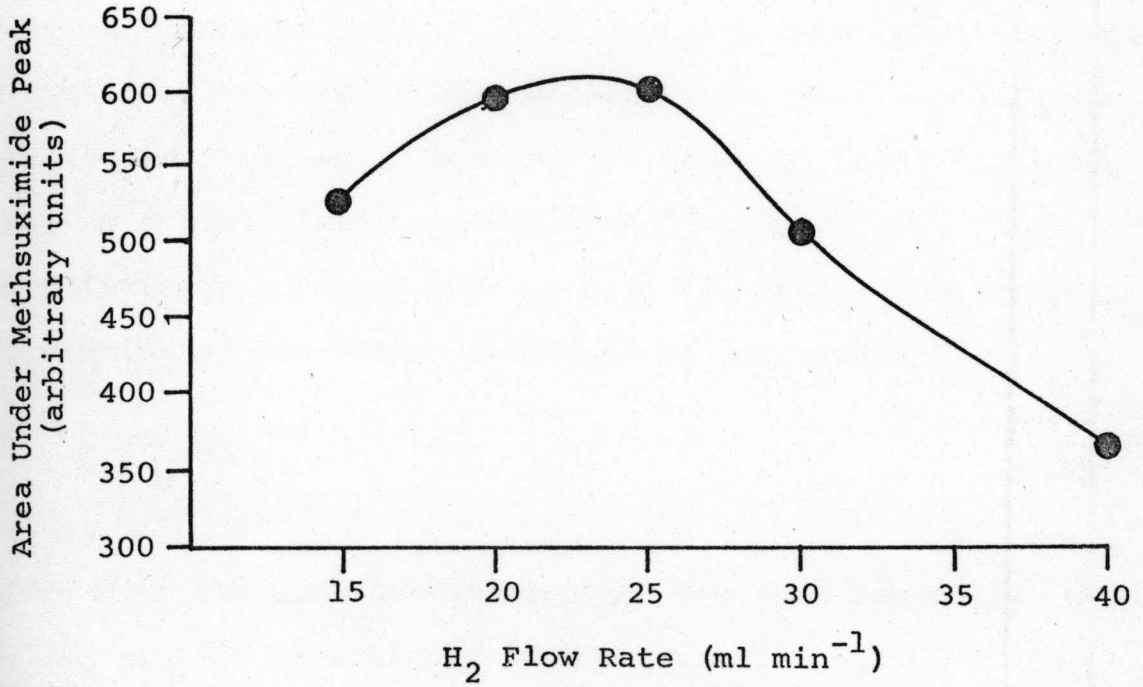
$$N = 16 \left( \frac{t_{\text{corr}}}{W} \right)^2 \quad \text{Eq. 1}$$

$$\bar{N} = \frac{N}{L} \quad \text{Eq. 2}$$

$$\text{HETP} = \frac{1}{\bar{N}} \quad \text{Eq. 3}$$

where N is the number of theoretical plates, t<sub>corr</sub> is the

Figure 2. Area under drug (upper) and metabolite (lower) GC peaks determined by disc integration versus H<sub>2</sub> flow rate. Air flow was 10 X the H<sub>2</sub> flow rate and column temperature 175° C with N<sub>2</sub> flow at 20 ml min<sup>-1</sup>. The points were experimentally determined and lines were hand drawn.



time between the air peak<sup>a</sup> and the compound's peak apex,  $W$  is the baseline width of the triangulated peak,  $t_{\text{corr}}$  and  $W$  have the same units,  $\bar{N}$  is  $N$  divided by column length in feet,  $L$ . HETP is usually expressed in centimeters. A larger  $\bar{N}$  indicates greater column efficiency for the compounds on the specific column. Reference 19 gives a rating scale for judging column efficiency based on  $\bar{N}$  values. Regardless of efficiency a column must be able to separate the compounds. Resolution of two peaks is defined by equation 4

$$R = \frac{2D}{W_1 + W_2} \quad \text{Eq. 4}$$

where  $D$  is the distance between the two peak apexes and  $W_1$  and  $W_2$  are the base widths of the two triangulated peaks. Generally, when two ideal peaks have an  $R = 1.5$  they overlap to the extent of about 0.3%. Table I indicates the resolution and efficiency of the column under the conditions of use. The Van Deemter plot is shown in Figure 3 and is the basis for the  $N_2$  flow rate of  $20 \text{ ml min}^{-1}$ .

### 3. Standard Curves

During the GC development over 300 peaks were quantitated by peak height and peak area determinations. Areas were calculated by triangulation, disc integration and planimetry. It was found that peak height ratios of drug or metabolite to phensuximide external standard were more

<sup>a</sup>Under the experimental conditions the time of the air peak was considered to be zero, the injection point.

Table IGC Column Efficiency Parameter Values

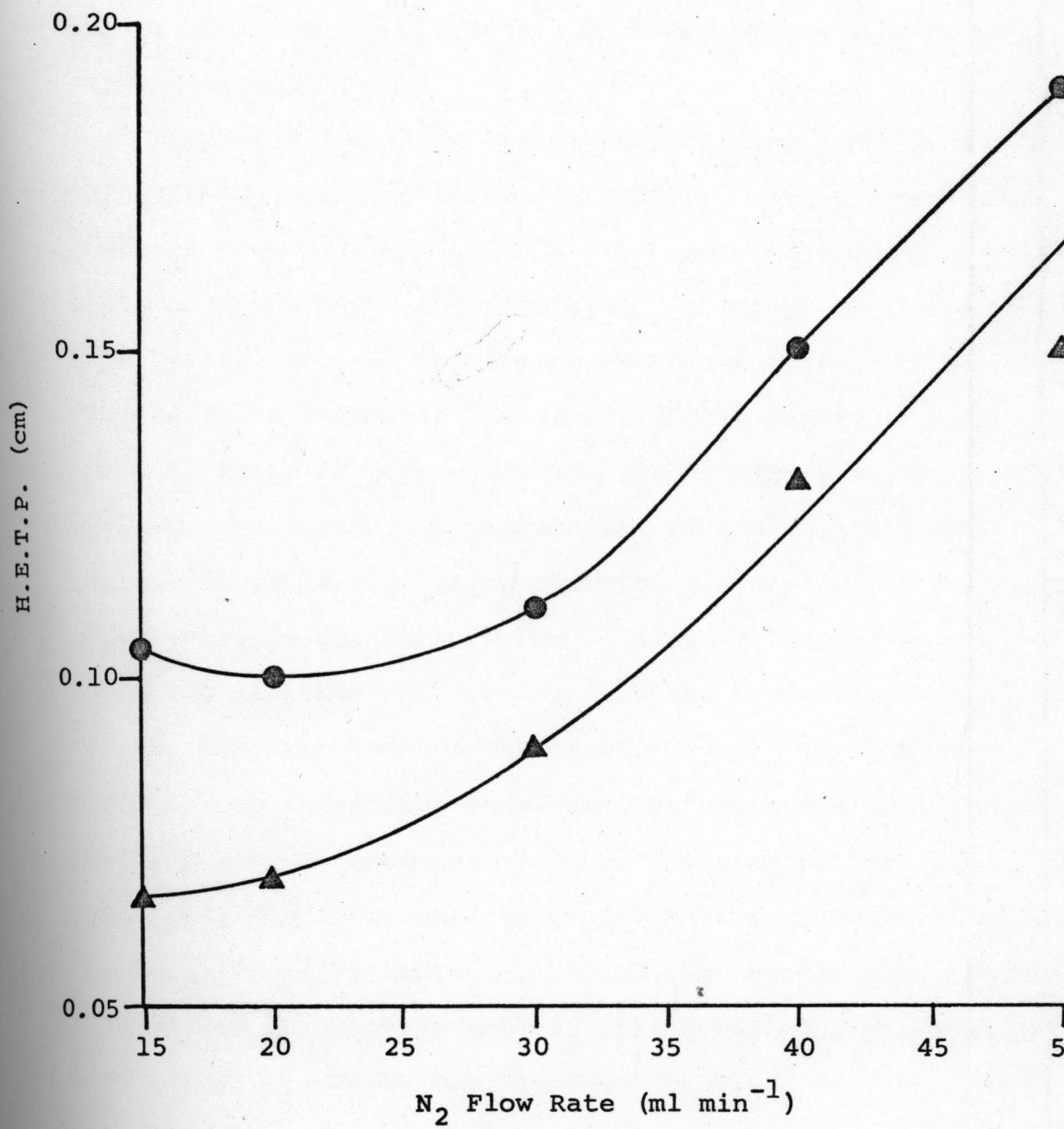
	<u>methsuximide</u>	<u>2-methyl-2-phenylsuccinimide</u>
HETP (cm)	0.095	0.070
$\bar{N}$ (ft <sup>-1</sup> )	321	435
R <sup>a</sup>	1.4	1.0
Rating <sup>b</sup>	good	good

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<sup>a</sup>Resolution is with respect to the nearest peak, phensuximide.

<sup>b</sup>See reference 19.

Figure 3. The Van Deemter plot; HETP (cm) vs  $N_2$  flow rate ( $ml\ min^{-1}$ ). Points were experimentally determined, methsuximide (●) and metabolite (▲), while lines were hand drawn.



reproducible than peak area ratios. Efforts were made to obtain sharp narrow peaks so that peak heights would be a reliable quantitative method. Area determination of thin peaks leads to considerable error by any method other than digital computation, since the thickness of the pen tracing becomes significant.

Figures 4 and 5 are the standard curves used to quantitate unknown plasma drug concentrations. Seven drug concentrations ranging from 0.5-15  $\mu\text{g ml}^{-1}$  were used. Triplicate runs were made using the same stock solutions so that three data points were obtained at each concentration. This allowed for a statistical analysis of the linear regression model in terms of pure error and lack of fit variance contributions. Pure error is a component of the variance not explained by the regression equation and includes errors of replication, i.e., assay errors. Lack of fit is the remaining variance not explained by the regression equation and can indicate lack of fit of the data to the proposed model. Both regression equation intercepts are not significantly different from zero so that the peak height ratios are directly proportional to concentrations. Table II gives the analysis of variance (ANOVA) of the regressions. Nomenclature and calculation methods were obtained from Draper and Smith (20). Although the "goodness of fit"<sup>a</sup>,  $R^2$ , and correlation coefficient,  $r$ , are excellent, Table II indicates that

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<sup>a</sup>Coefficient of determination,  $R^2$ , is  $\frac{\sum \text{obs.}^2 - \sum \text{dev.}^2}{\sum \text{obs.}^2}$ .

Figure 4. Methsuximide standard curve, peak height ratio (methsuximide:phensuximide) versus plasma concentration ( $\mu\text{g ml}^{-1}$ ). Regression line is  $y = 0.16 (\pm 0.01)X + 0.00 (\pm 0.08)$  where quantities in parenthesis are the 95% confidence limits.  $R^2 = 0.9795$  and  $r = 0.9995$ . Open circles indicate one determination, half-filled circles and filled circles indicate two and three equivalent determinations, respectively.

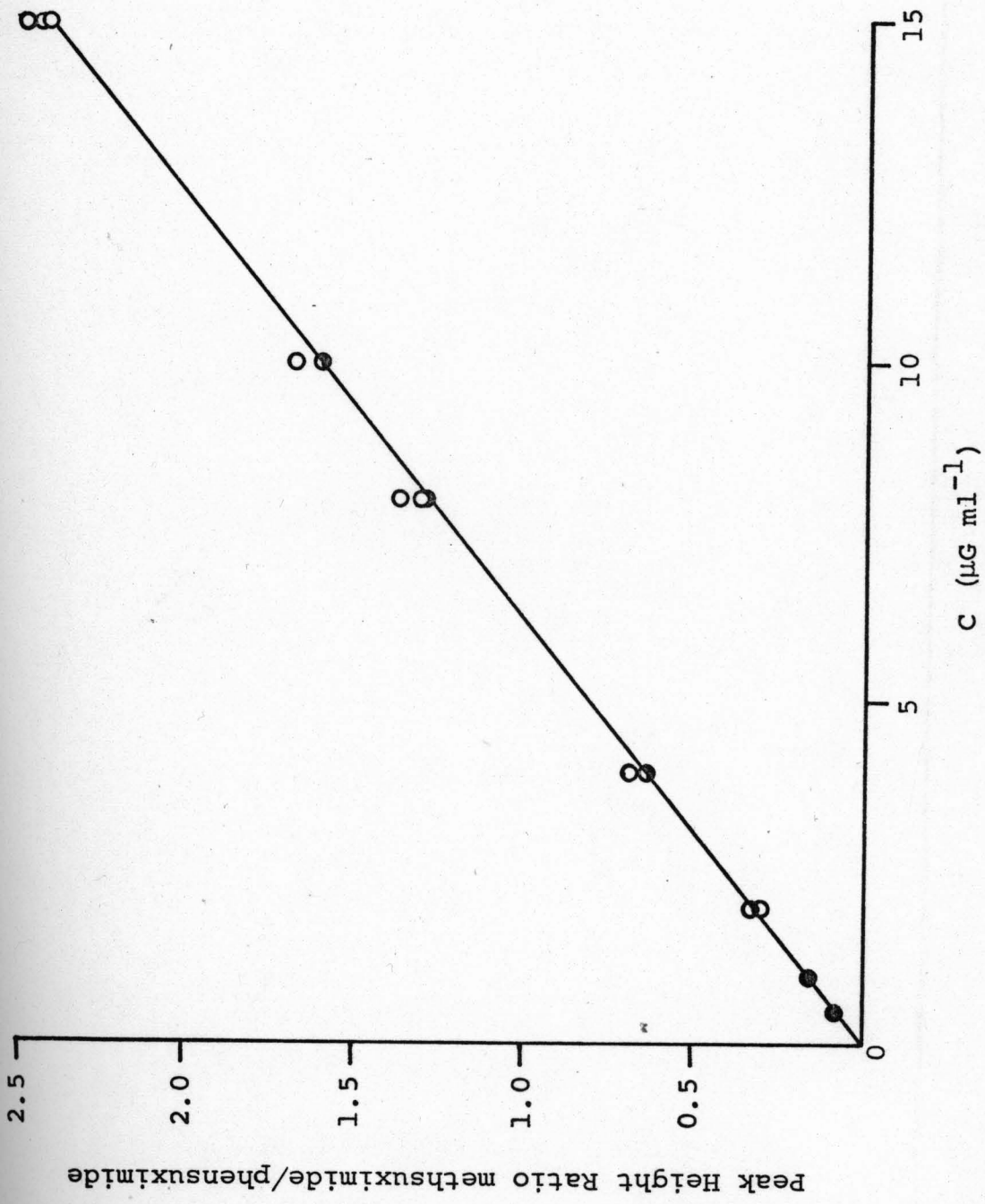


Figure 5. 2-Methyl-2-phenylsuccinimide standard curve, peak height ratio (metabolite:phensuximide) versus plasma concentration ( $\mu\text{g ml}^{-1}$ ). Regression line is  $y = 0.11 (\pm 0.004)X - 0.02 (\pm 0.029)$  where quantities in parenthesis are 95% confidence limits.  $R^2 = 0.9949$  and  $r = 0.9985$ . Symbols are as in Figure 4.

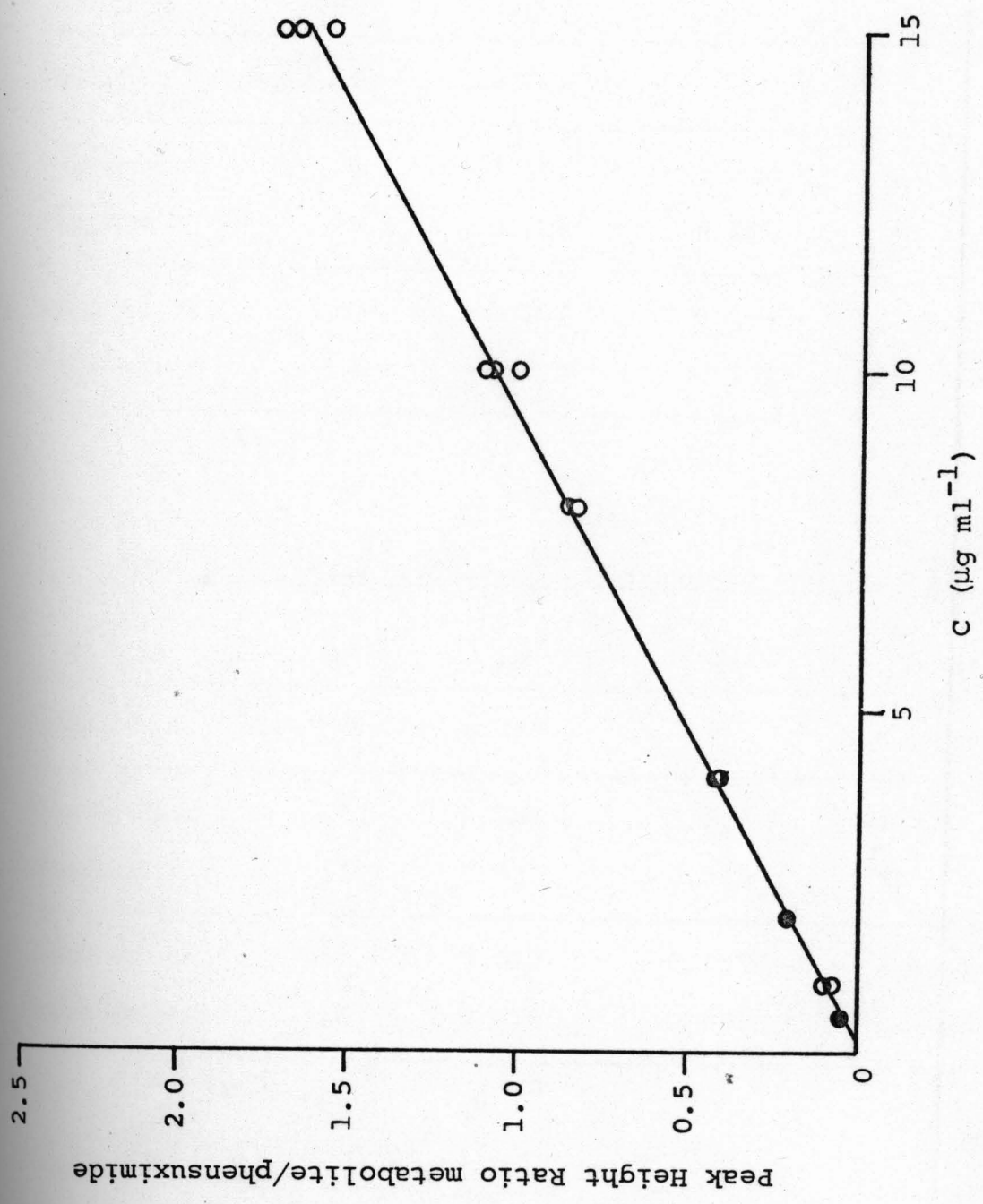


Table IIANOVA for Methsuximide Standard Curve

Source	df	SS	MS	F ratio
Total	20	14.068		
Regression	1	13.780	13.780	1005.8
Residual	19	0.288	0.014	(P < 0.01)
Lack of fit	5	0.276	0.055	55
Pure error	14	0.012	0.001	(P < 0.01)

ANOVA for 2-Methyl-2-phenylsuccinimide Standard Curve

Source	df	SS	MS	F ratio
Total	20	6.318		
Regression	1	6.285	6.285	3142
Residual	19	0.033	0.002	(P < 0.01)
Lack of fit	5	0.017	0.003	3 borderline
Pure error	14	0.016	0.001	(P = 0.05)

the methsuximide linear model suffers from a significant lack of fit. This problem was not investigated further as the linear model was judged sufficient to meet the demands of these experiments. Caution was exercised in extrapolations of the regression equation beyond  $15 \mu\text{g ml}^{-1}$ . In cases where plasma levels exceeded  $15 \mu\text{g ml}^{-1}$  the linear model was checked by spiking blank plasma at the higher concentrations. Deviations from the concentrations predicted by the linear model did not exceed  $\pm 5\%$ .

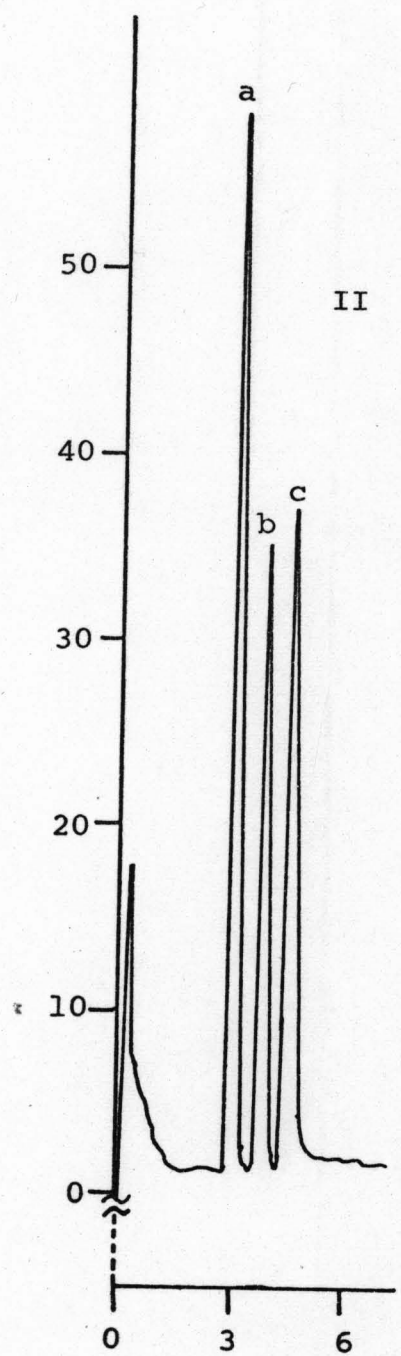
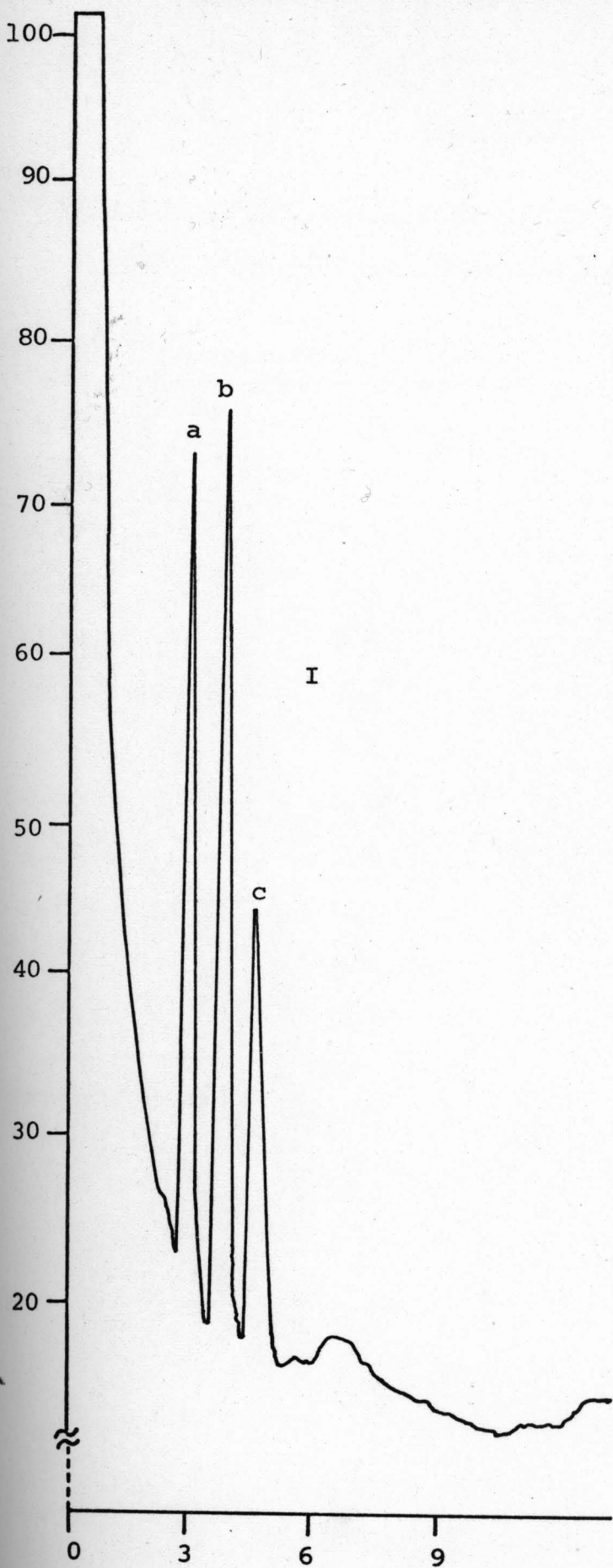
#### 4. Representative GC Chart

Representative GC charts with  $\text{CHCl}_3$  and  $\text{CS}_2$  as solvents are given in Figure 6. Retention times were methsuximide 191 sec, phensuximide external standard 239 sec and metabolite 281 sec with the  $\text{CS}_2$  system.

#### B. Intravenous Drug and Metabolite Plasma Level-Time Data

Plasma levels of drug and metabolite after i.v. doses of methsuximide are given in Tables III-V and graphically in Figures 7-9. Plasma levels of metabolite after dosing this compound are described in Tables VI and VII and Figures 10 and 11. The solid lines in the figures are best computer fits as described in the text.

Figure 6. Representative GC chart. Solvent peaks in I and II are  $\text{CHCl}_3$  and  $\text{CS}_2$  respectively. Peaks a, b and c are methsuximide, phensuximide and 2-methyl-2-phenylsuccinimide, respectively.



TIME (minutes)

Table III

Drug and Metabolite Plasma Levels after Intravenous  
Methsuximide Administration to Dog A, Experiment A

<u>t (hrs)</u>	<u>C<sub>methsuximide</sub> (<math>\mu\text{g ml}^{-1}</math>)</u>	<u>C<sub>metabolite</sub> (<math>\mu\text{g ml}^{-1}</math>)</u>
0.16	13.6	3.5
0.27	12.2	4.3
0.50	7.2	5.0
0.75	5.6	5.5
1.00	5.6	5.5
2.25	--- <sup>a</sup>	7.0
3.00	4.4	7.0
4.00	3.4	6.5
6.00	1.6	6.5
8.00	1.3	5.5
10.00	0.9	5.0
12.00	0.7	4.5

<sup>a</sup>Not determined.

Table IV

Drug and Metabolite Plasma Levels after Intravenous  
Methsuximide Administration to Dog B, Experiment B

<u>t (hrs)</u>	<u>C<sub>methsuximide</sub> (<math>\mu\text{g ml}^{-1}</math>)</u>	<u>C<sub>metabolite</sub> (<math>\mu\text{g ml}^{-1}</math>)</u>
0.08	12.6	--- <sup>a</sup>
0.17	9.1	---
0.25	6.9	1.8
0.50	5.7	3.3
0.75	4.0	4.5
1.00	3.0	4.9
2.00	1.6	6.1
3.00	0.8	6.2
4.00	0.5	5.9
6.00	--- <sup>a</sup>	5.2
8.00	---	4.9
10.00	---	4.8
12.00	---	4.9
14.00	---	4.7
16.00	---	3.9
18.00	---	3.7

<sup>a</sup>Below limits of detection.

Table V

Drug and Metabolite Plasma Levels after Intravenous  
Methsuximide Administration to Dog B, Experiment C

<u>t (hrs)</u>	<u>C<sub>methsuximide</sub> (<math>\mu\text{g ml}^{-1}</math>)</u>	<u>C<sub>metabolite</sub> (<math>\mu\text{g ml}^{-1}</math>)</u>
0.03	10.7	--- <sup>a</sup>
0.08	8.6	0.6
0.25	6.7	1.4
0.50	5.3	2.0
0.75	4.7	2.9
1.00	3.9	2.6
1.50	2.9	3.4
2.00	1.8	3.3
3.00	1.3	4.4
4.00	0.9	4.5
5.00	0.6	4.2
6.00	0.4	4.5
8.00	--- <sup>a</sup>	4.3
10.00	---	4.4
12.00	---	4.4
24.00	---	2.4
36.00	---	1.3
48.00	---	0.5

<sup>a</sup>Below limits of detection.

Table VI

Metabolite Plasma Levels after Intravenous 2-Methyl-2-phenylsuccinimide Administration to Dog B, Experiment D

<u>t (hrs)</u>	<u>C<sub>metabolite</sub> (<math>\mu\text{g ml}^{-1}</math>)</u>
0.09	19.8
0.17	15.0
0.25	18.0
0.50	16.4
0.75	14.8
1.00	16.5
2.00	13.2
3.20	14.1
4.00	16.1
6.00	12.2
8.00	12.6
10.00	11.4
12.00	9.1
18.00	7.8
24.00	5.7
36.00	3.4
48.00	1.6

Table VII

Metabolite Plasma Levels after Intravenous 2-Methyl-2-phenylsuccinimide Administration to Dog B, Experiment E

<u>t (hrs)</u>	<u>C<sub>metabolite</sub> (μg ml<sup>-1</sup>)</u>
0.08	24.0
0.17	21.3
0.25	19.0
0.50	19.2
1.00	17.9
2.00	17.5
3.80	17.9
6.00	17.4
8.00	14.5
11.00	11.7
13.00	12.2
18.00	10.4
24.00	7.5
36.00	4.3
48.00	1.9

Figure 7. Methsuximide and 2-methyl-2-phenylsuccinimide plasma level vs time profiles after a 250 mg intravenous injection of methsuximide, experiment A. The methsuximide profile (●) was drawn using the computer determined equation  $C_1 = 16.7e^{-4.71t} + 6.6e^{-0.20t}$  while the metabolite profile (▲) was hand drawn.

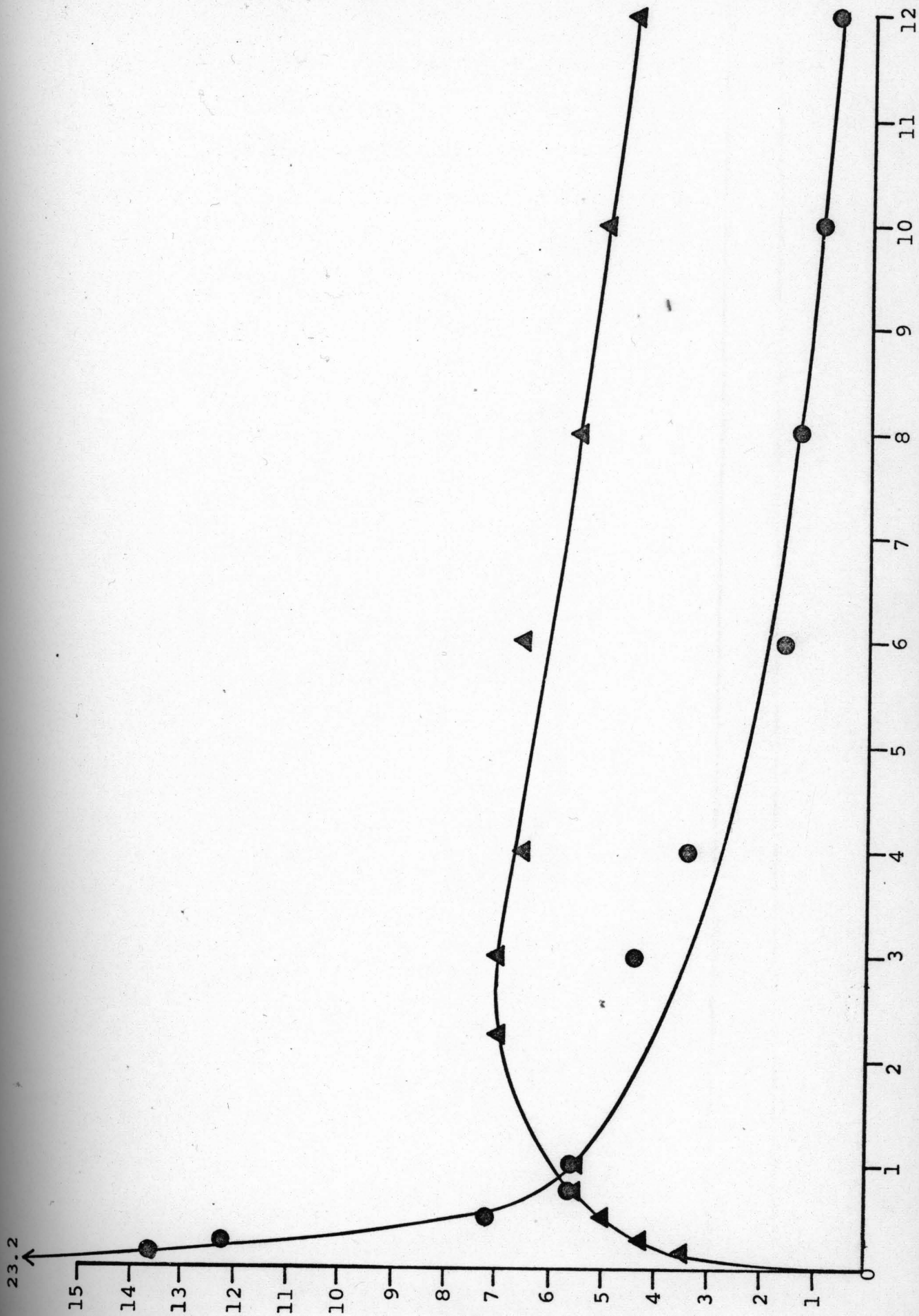


Figure 8. Methsuximide and 2-methyl-2-phenylsuccinimide plasma level vs time profiles after a 250 mg intravenous injection of methsuximide, experiment B. Computer determined equations are methsuximide (●)

$$C_1 = 11.6e^{-8.64t} + 7.2e^{-0.78t}$$

and metabolite (▲)

$$C_{M_b} = -1.0e^{-8.64t} - 7.1e^{-0.78t} + 8.1e^{-0.047t}$$

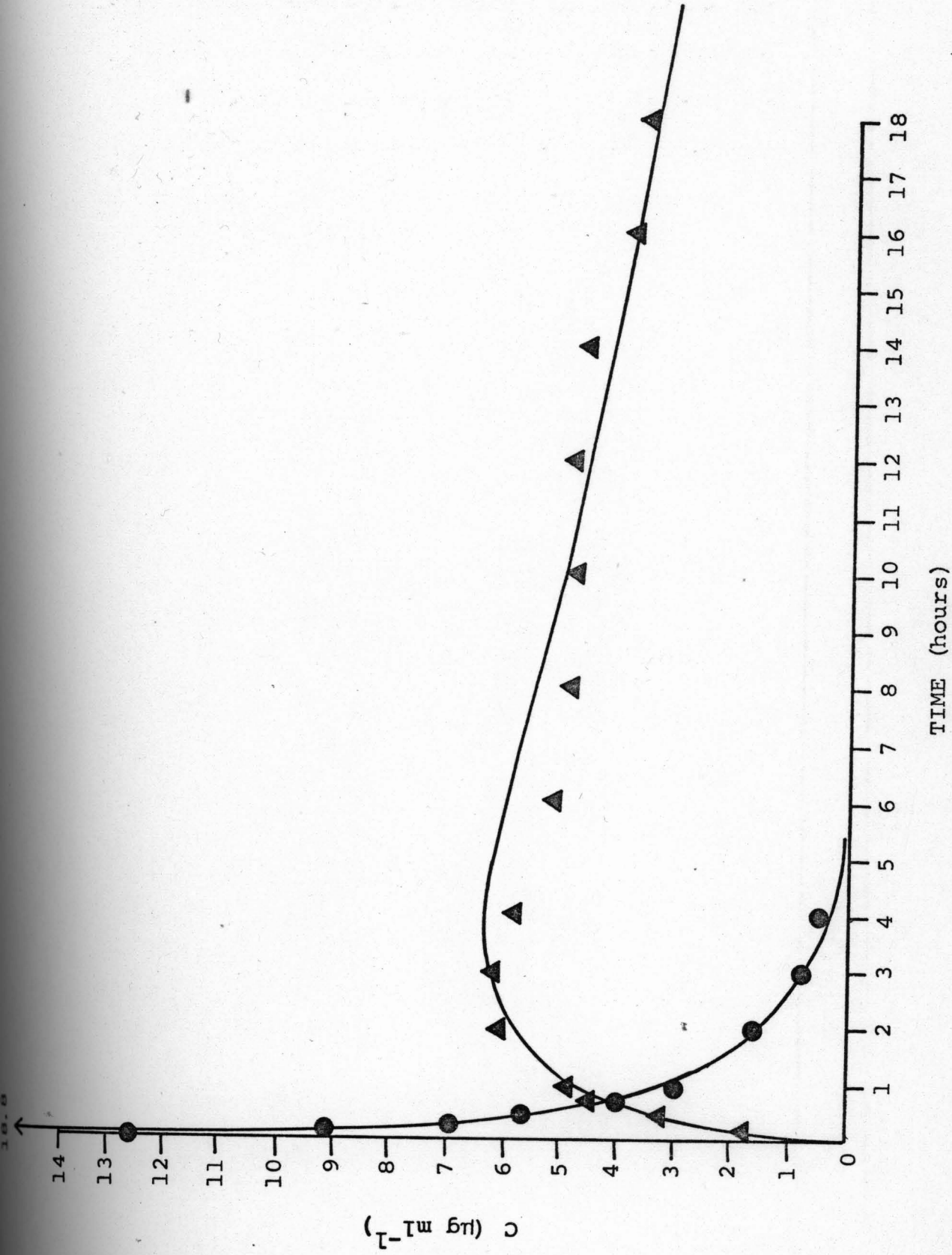


Figure 9. Methsuximide and 2-methyl-2-phenylsuccinimide plasma level vs time profiles after a 250 mg intravenous injection of methsuximide, experiment C. Computer determined equations are

methsuximide (●)

$$C_1 = 6.2e^{-2.24t} + 4.2e^{-0.40t}$$

and metabolite (▲)

$$C_{M_b} = -1.4e^{-2.24t} - 5.6e^{-0.40t} + 7.0e^{-0.047t}$$

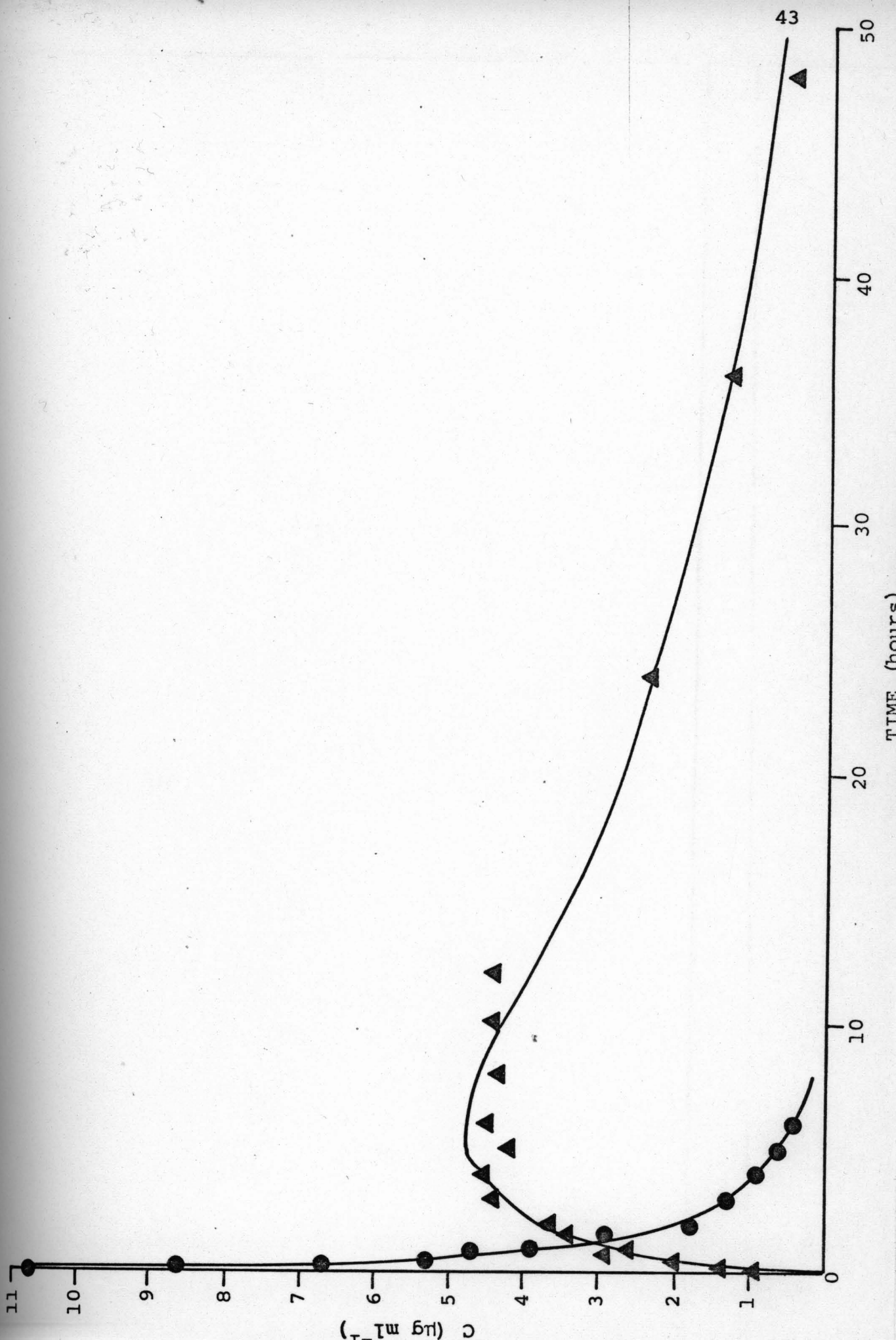


Figure 10. 2-Methyl-2-phenylsuccinimide plasma level vs time profile after a 250 mg injection of this metabolite, experiment D. The equation describing the profile is  $C_{M_b} = 17.1e^{-0.047t}$ .

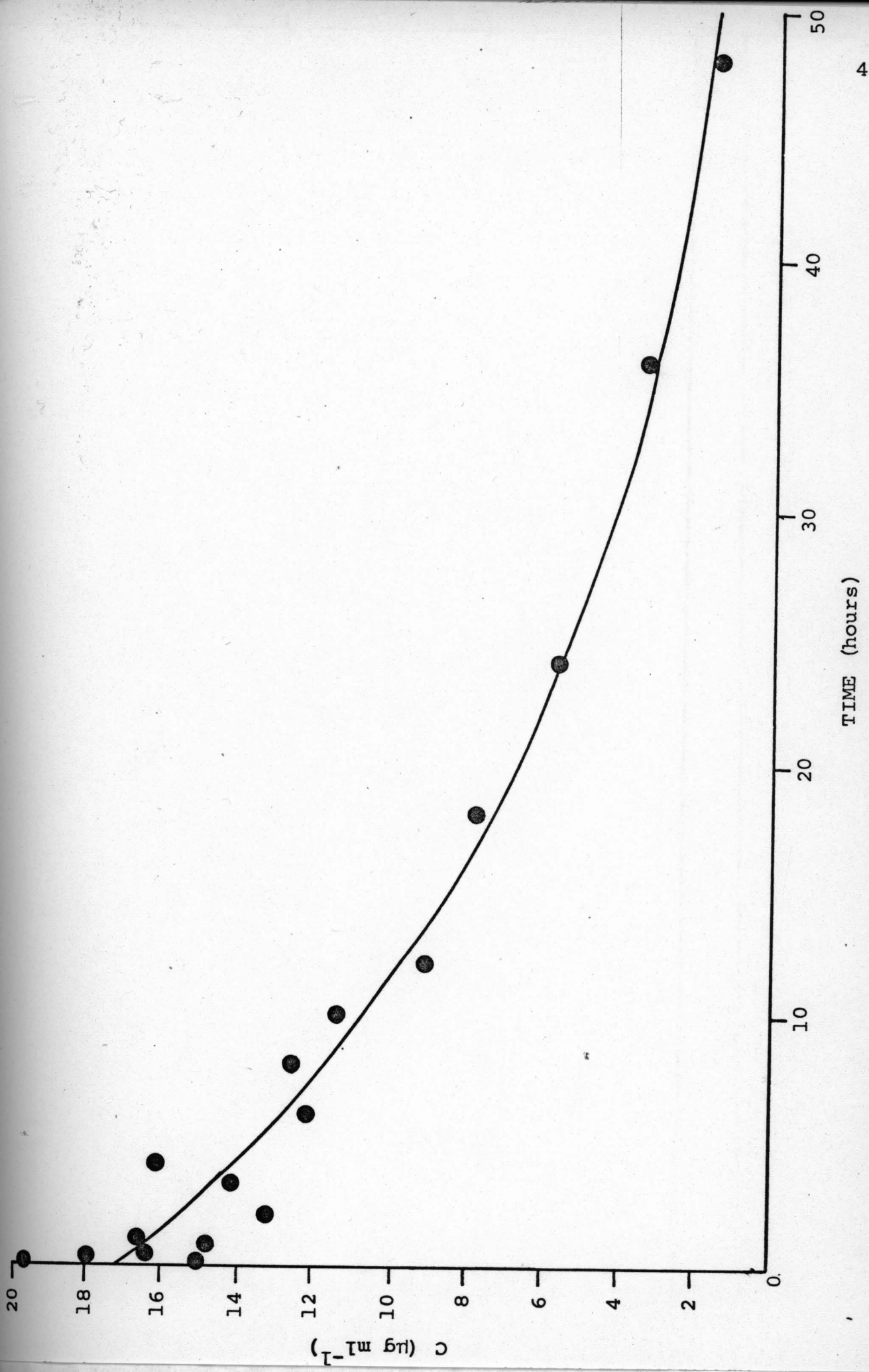
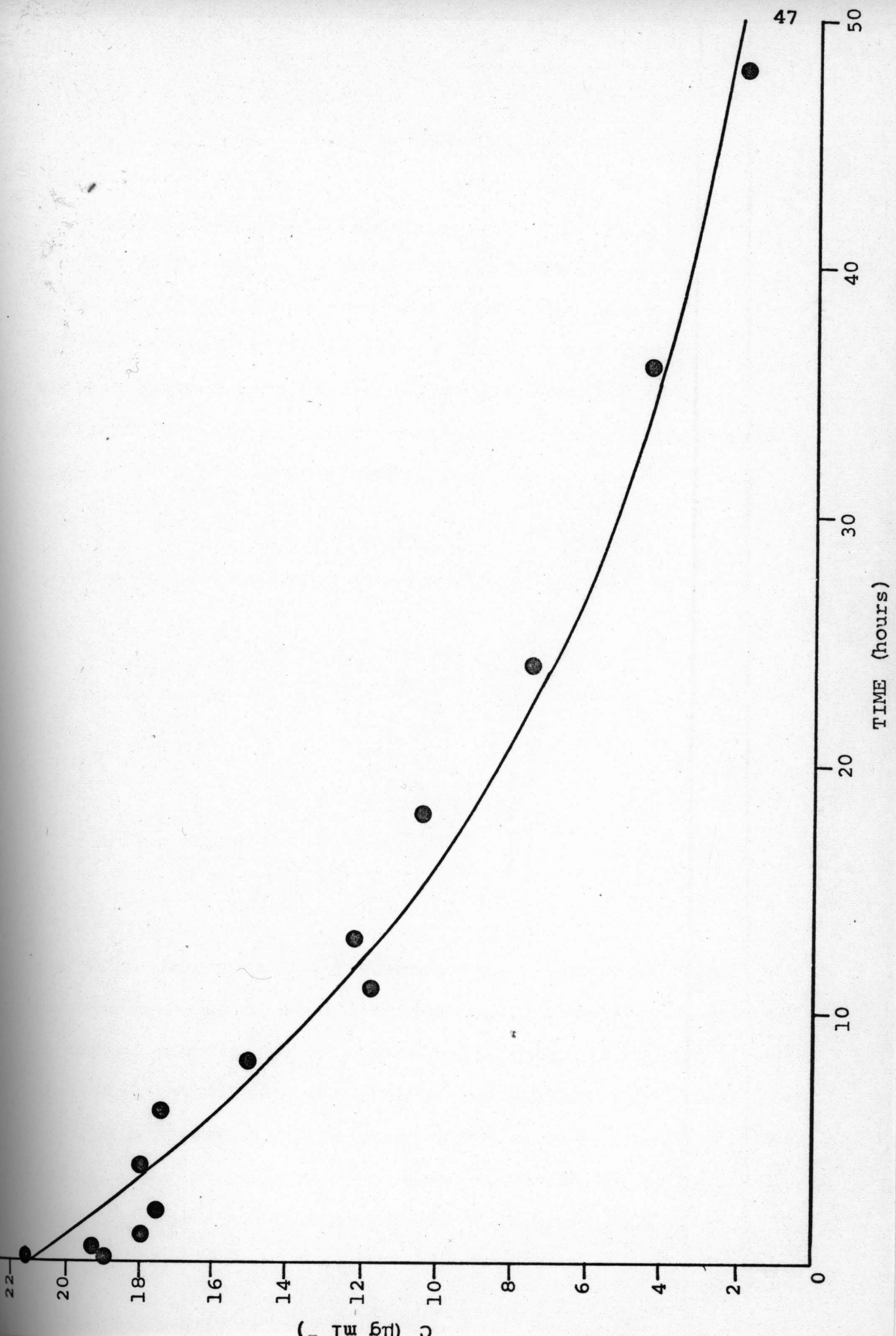


Figure 11. 2-Methyl-2-phenylsuccinimide plasma level vs time profile after a 250 mg injection of this metabolite, experiment E. The equation describing the profile is  $C_{M_b} = 21.1e^{-0.047t}$ .



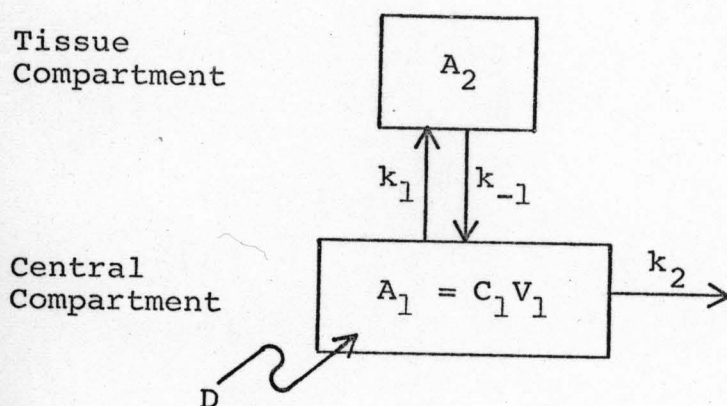
## DISCUSSION

A. Methsuximide Pharmacokinetics

The methsuximide concentration vs time profiles are given in Figures 12-14 on a semilogarithmic scale. The figures indicate that the plasma level versus time profile after intravenous methsuximide is biexponential and may be analyzed in terms of the two compartment open pharmacokinetic model (20) depicted in Scheme I.

Scheme I

Two Compartment Open Model with Intravenous Injection



The model describes the instantaneous introduction of an intravenous dose of drug  $D$  into a rapidly accessible central or plasma compartment of apparent distribution volume  $V_1$  and equilibration between the central compartment and a less readily accessible tissue compartment  $A_2$ . The quantities  $A_1$  and  $A_2$  refer to amounts of unchanged drug in the central and tissue compartments respectively. The disposition first

Figure 12. Semilogarithmic plot of drug concentration-time data of Table III showing biexponential decline in methsuximide plasma level. Equations are computer fits to Scheme I model using unweighted data (- - -) and data weighted by  $1/C$  (—). The equations are  $C_1 = 15.4e^{-4.01t} + 6.2e^{-0.18t}$  and  $C_1 = 16.7e^{-4.71t} + 6.6e^{-0.20t}$  respectively.

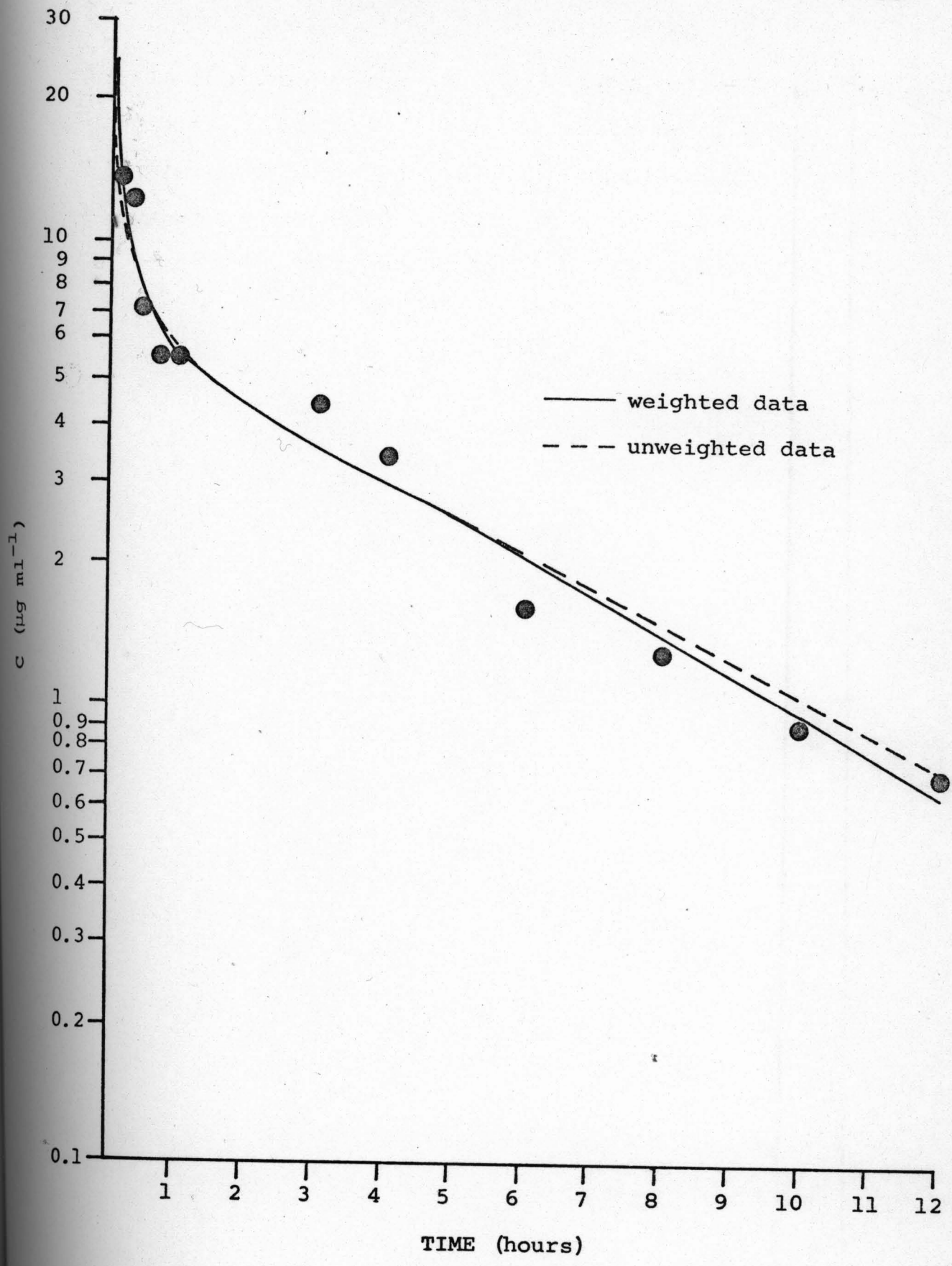


Figure 13. Semilogarithmic plot of drug concentration-time data of Table IV showing biexponential decline in methsuximide plasma level. Equations are computer fits to Scheme I model using unweighted data (- - -) and data weighted by  $1/C$  (——). The equations are  $C_1 = 12.1e^{-9.95t} + 7.7e^{-0.81t}$  and  $C_1 = 11.7e^{-8.03t} + 6.8e^{-0.71t}$  respectively.

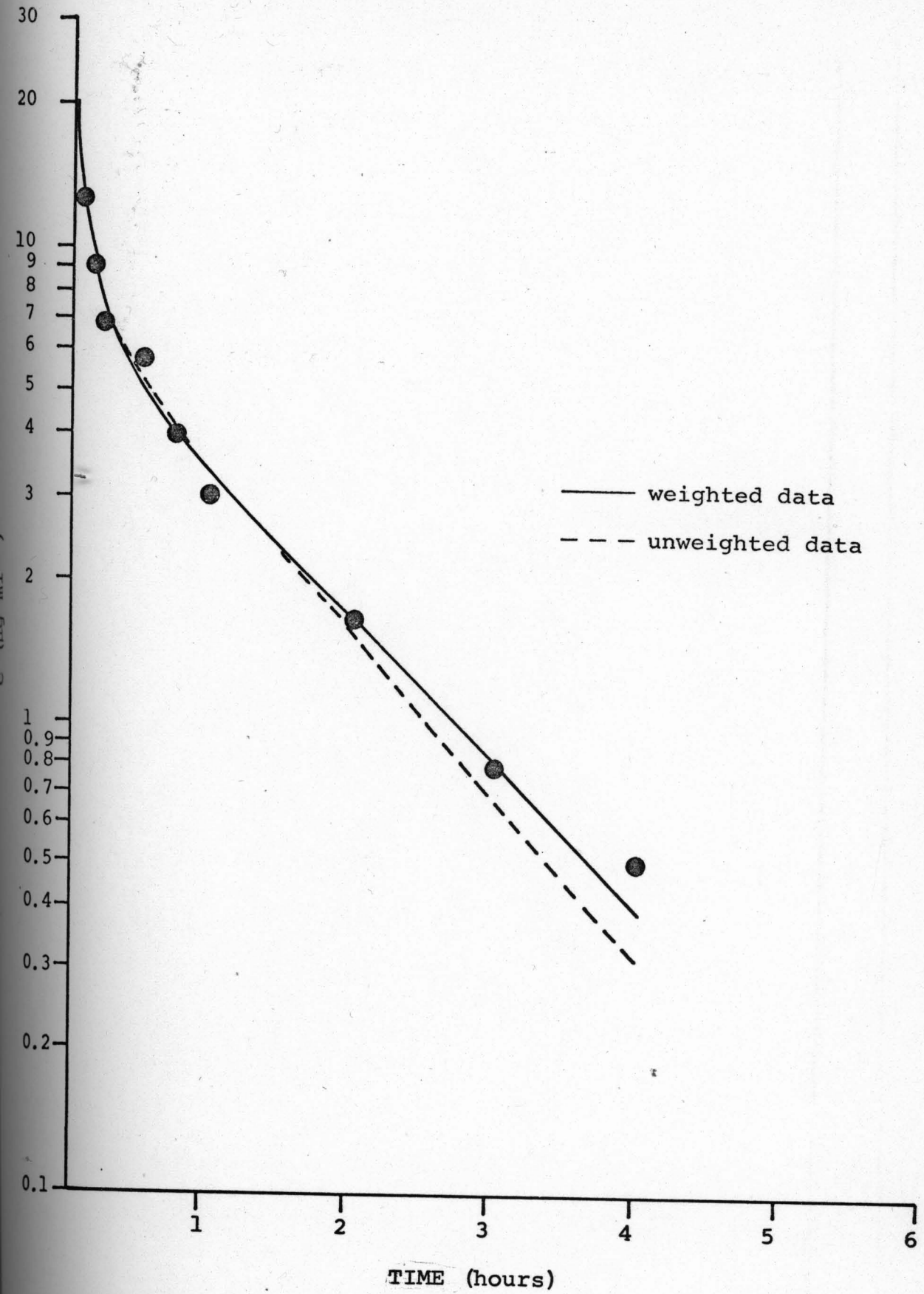
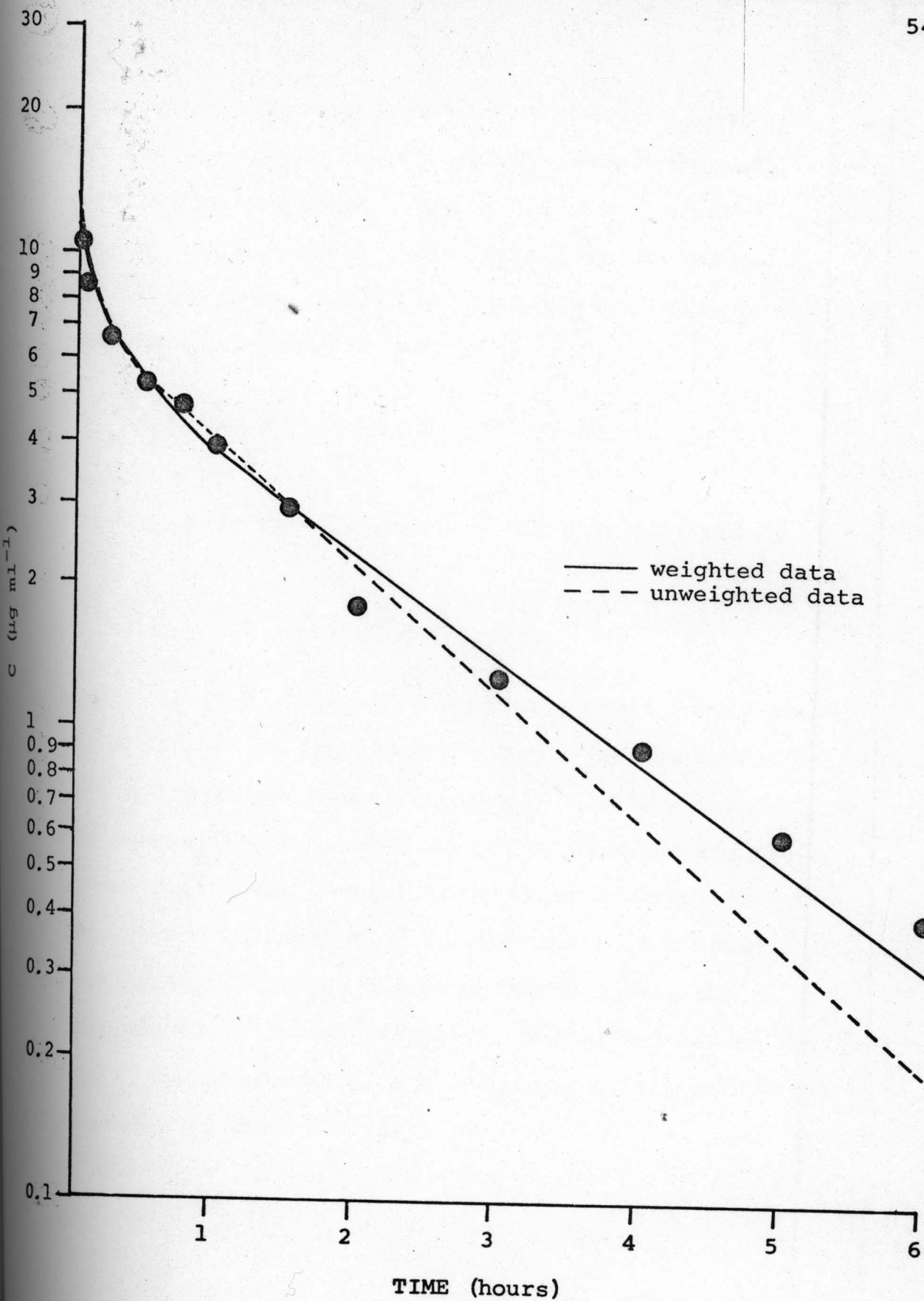


Figure 14. Semilogarithmic plot of drug concentration-time data of Table V showing biexponential decline in methsuximide plasma level. Equations are computer fits to Scheme I model using unweighted data (- - -) and data weighted by  $1/C$  (—). The equations are  $C_1 = 5.3e^{-14.26t} + 7.3e^{-0.61t}$  and  $C_1 = 4.9e^{-5.48t} + 6.2e^{-0.50t}$  respectively.



order rate constants  $k_1$  and  $k_{-1}$  govern the transfer of drug between the compartments while  $k_2$  is the first order rate constant representing loss of drug from the central compartment by all processes, i.e., excretion and metabolism.

Equation 5 describes the drug plasma concentration vs time profile appropriate to this model (21).

$$C_1 = \frac{D}{V_1(\alpha - \beta)} [(\alpha - k_{-1})e^{-\alpha t} + (k_{-1} - \beta)e^{-\beta t}] \quad \text{Eq. 5}$$

The composite rate constants  $\alpha$  and  $\beta$  are defined by

$$\alpha, \beta = \frac{1}{2} [(k_1 + k_{-1} + k_2) \pm \sqrt{(k_1 + k_{-1} + k_2)^2 - 4k_{-1}k_2}] \quad \text{Eq. 6}$$

where  $\beta$  is the slope of the post-distributive log-linear phase of loss of drug from the body, divided by 2.303, and  $\alpha$  is the log-linear slope, divided by 2.303, obtained from subtracting the extrapolated  $\beta$  line from the actual data points during the initial curvilinear distributive phase. This method of separating exponential rate constants has been described in the pharmacokinetic literature as 'feathering', 'curve stripping', or 'method of residuals' (21). The rate constants  $k_1$ ,  $k_{-1}$  and  $k_2$  are determined through equations 7-9 (21).

$$k_{-1} = \frac{A\beta + B\alpha}{A + B} \quad \text{Eq. 7}$$

$$k_2 = \frac{\alpha\beta}{k_{-1}} \quad \text{Eq. 8}$$

$$k_1 = \alpha + \beta - k_{-1} - k_2 \quad \text{Eq. 9}$$

where A and B are the intercepts on the concentration axis of the  $\alpha$  and  $\beta$  extrapolated lines, respectively. The apparent volume of distribution  $V_1$  is determined with intravenous data by,

$$V_1 = \frac{D}{A + B} \quad \text{Eq. 10}$$

The biological half-life  $t_{1/2}$  is determined by equation 11 while T/P, the ratio of amount of drug in the tissue compartment to that in the central compartment at equilibrium, is determined by equation 12 (21).

$$t_{1/2} = \frac{0.693}{\beta} \quad \text{Eq. 11}$$

$$T/P = \frac{k_1}{k_{-1} - \beta} \quad \text{Eq. 12}$$

Subsequent to graphical analysis, improved estimates of all parameters together with confidence limits, correlation coefficients and coefficients of determination were obtained using the iterative nonlinear regression program NREG<sup>a</sup> on a digital computer<sup>b</sup>. Input into the computer consisted of graphical estimates of parameters together with the relevant

<sup>a</sup>With user supplied subroutine - see Appendix A for a typical NREG output.

<sup>b</sup>Univac 1110 digital computer, Madison Academic Computing Center, the University of Wisconsin, Madison.

equations contained in the user supplied subroutine. Table VIII gives the pharmacokinetic parameters describing the methsuximide kinetics after intravenous administration based on the computer estimates. The dashed lines in Figures 12-14 are based on these values. Computer fits of the terminal data were poor and this is especially evident in experiment C. Since NREG is a least squares fitting routine a small relative error in the higher concentration range will contribute more to the sums of squares than an equivalent error in the lower concentration range. In order to make all data points contribute equally to the sums of squares, concentrations were individually weighted by the factor  $1/C$ . The computer estimated parameters with weighted data are given in Table IX while the computer fits are the solid lines in Figures 12-14. The fits resulting from the weighted data were judged better than those from the unweighted data on the basis of the overall statistical computer outputs (sums of squares, correlation matrix, confidence limits on linear hypothesis, exploration, graph of residuals,  $R^2$ ,  $r$ , etc. - see Appendix A). Table IX indicates that the distribution characteristics of methsuximide,  $k_1$ ,  $k_{-1}$ ,  $\alpha$ ,  $V_1$  and T/P, differed between the two dogs although they were similar orders of magnitude. There was more rapid equilibration of drug between the two compartments in Dog B ( $\alpha$ ,  $k_1$ ,  $k_{-1}$ ) while Dog A had a greater amount of drug in the tissue compartment at equilibrium than Dog B (T/P). The elimination characteristics,  $k_2$ ,  $\beta$  and  $t_{1/2}$ , also differed between the two dogs. Dog A eliminated

Methsuximide Computer Estimated Pharmacokinetic Parameters Using Unweighted Data

	Experiment A, Dog A	Experiment B, Dog B	Experiment C, Dog B
D mg	250	250	250
$V_1$ l	11.6	12.6	19.8
$D/V_1$ mg l <sup>-1</sup>	21.5 (15.0-28.1) <sup>a</sup>	19.8 (15.0-24.5)	12.6 (11.2-14.1)
A $\mu$ g ml <sup>-1</sup>	15.4	12.2	5.3
B $\mu$ g ml <sup>-1</sup>	6.2	7.7	7.4
$\alpha$ hr <sup>-1</sup>	4.01	9.95	14.26
$\beta$ hr <sup>-1</sup>	0.18	0.81	0.61
$t_{1/2}$ hrs	3.85	0.85	1.14
$k_1$ hr <sup>-1</sup>	2.34 (0.64-4.05)	4.55 (1.53-7.57)	5.29 (1.96-8.63)
$k_{-1}$ hr <sup>-1</sup>	1.28 (0.43-2.13)	4.35 (2.08-6.62)	8.55 (4.70-12.40)
$k_2$ hr <sup>-1</sup>	0.56 (0.33-0.79)	1.86 (1.30-2.42)	1.02 (0.87-1.17)
T/P	2.13	1.28	0.67
$R^2$	0.9935	0.9980	0.9987
r	0.9919	0.9975	0.9985

<sup>a</sup> 95% confidence intervals.

## Methsuximide Computer Estimated Pharmacokinetic Parameters

Where Each Concentration was Weighted by 1/Concentration

	Experiment A, Dog A	Experiment B, Dog B	Experiment C, Dog B
D mg	250	250	250
$V_1$ l	10.7	13.5	22.5
$D/V_1$ mg l <sup>-1</sup>	23.3 (12.9-33.7)	18.5 (13.4-23.6)	11.1 (9.6-12.6)
A $\mu\text{g ml}^{-1}$	16.7	11.7	4.9
B $\mu\text{g ml}^{-1}$	6.6	6.8	6.2
$\alpha$ hr <sup>-1</sup>	4.71	8.03	5.48
$\beta$ hr <sup>-1</sup>	0.20	0.71	0.50
$t_{1/2}$ hrs	3.47	0.98	1.39
$k_1$ hr <sup>-1</sup>	2.80 (0.32-5.29)	3.67 (1.02-6.32)	1.86 (0.074-3.65)
$k_{-1}$ hr <sup>-1</sup>	1.47 (0.69-2.26)	3.40 (1.89-4.91)	3.28 (0.73-5.82)
$k_2$ hr <sup>-1</sup>	0.63 (0.35-0.91)	1.67 (1.21-2.13)	0.84 (0.70-0.97)
T/P	2.20	1.36	0.67
$R_2$	0.9930	0.9977	0.9972
r	0.9913	0.9970	0.9964

and/or metabolized methsuximide more slowly. Differences were observed in parameters from experiments B and C in the same dog. This was not unexpected since the time period between experiments exceeded two months. For this reason each experiment was considered separately.

The rate constant  $k_2$  reflects excretion of unchanged methsuximide by any route and biotransformation to any metabolite. Since the 2-methyl-2-phenylsuccinimide was specifically quantitated in all methsuximide experiments it was possible to separate  $k_2$  into components representing 1) formation of 2-methyl-2-phenylsuccinimide and 2) elimination of unchanged methsuximide and formation of any other metabolites. A previous report (11) indicated that no unchanged methsuximide was found in 48 hour dog urine so that  $k_2$  primarily reflects biotransformation of methsuximide. Elimination by routes other than into urine (saliva, bile, etc.) is also possible. Before the 2-methyl-2-phenylsuccinimide data could be incorporated into Scheme I its apparent distribution volume  $V_3$  and individual pharmacokinetic model had to be obtained. This was accomplished by intravenous dosing of pure 2-methyl-2-phenylsuccinimide.

#### B. 2-Methyl-2-phenylsuccinimide (Metabolite) Pharmacokinetics

Figures 15 and 16 are semilogarithmic plots of data sets D and E (Tables VI-VII) obtained from intravenous metabolite experiments in Dog B on two separate occasions. Multiple blood samples during the first hour were necessary to

Figure 15. Semilogarithmic plot of metabolite concentration-time data of Table VI showing monoexponential decline in metabolite plasma level in dog B after dosing this compound. The linear regression equation including the 95% confidence limits on the slope and intercept is  $\log C = -0.0205 (\pm 0.0015)t + 1.2326 (\pm 0.0261)$ .

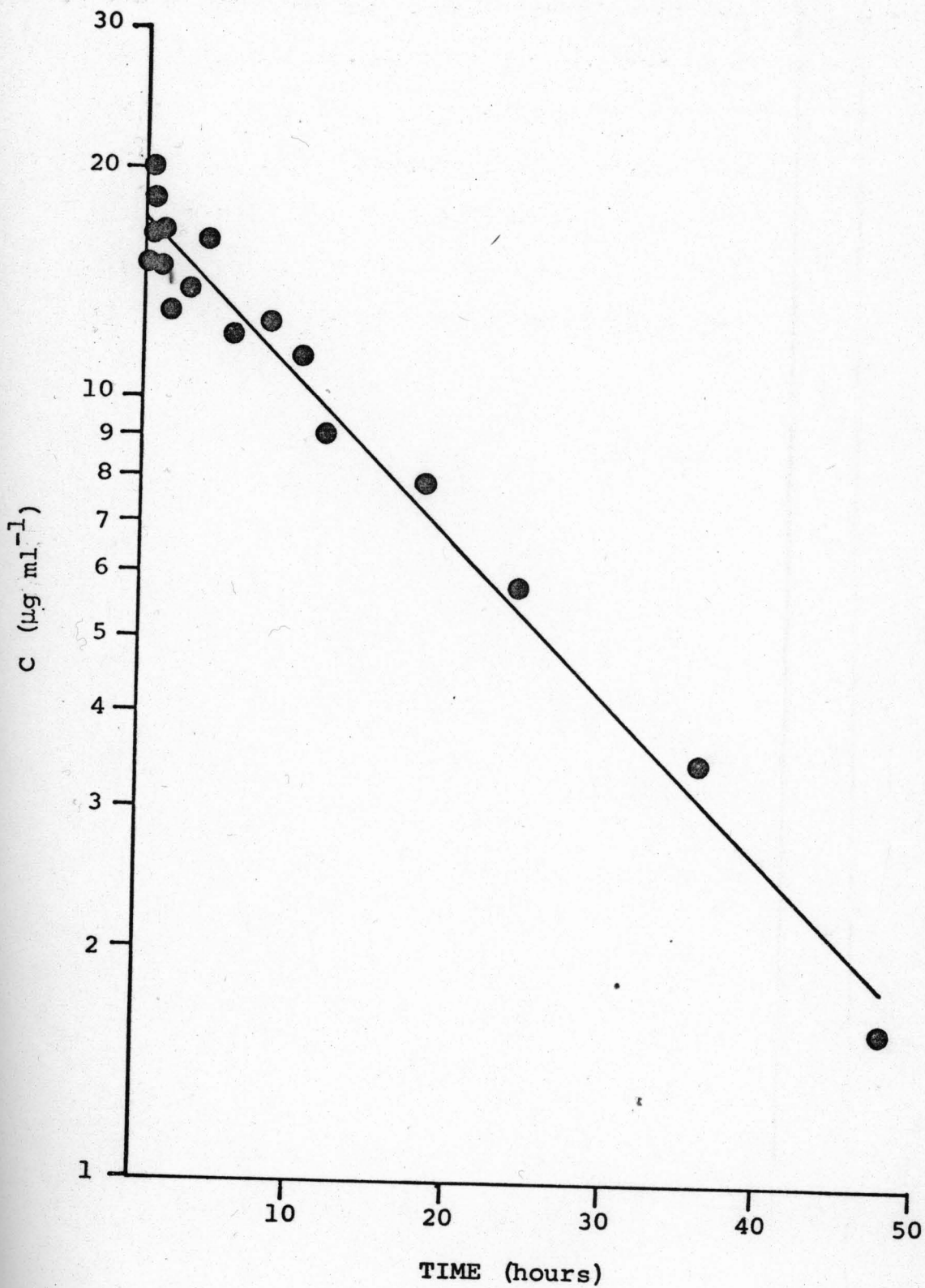
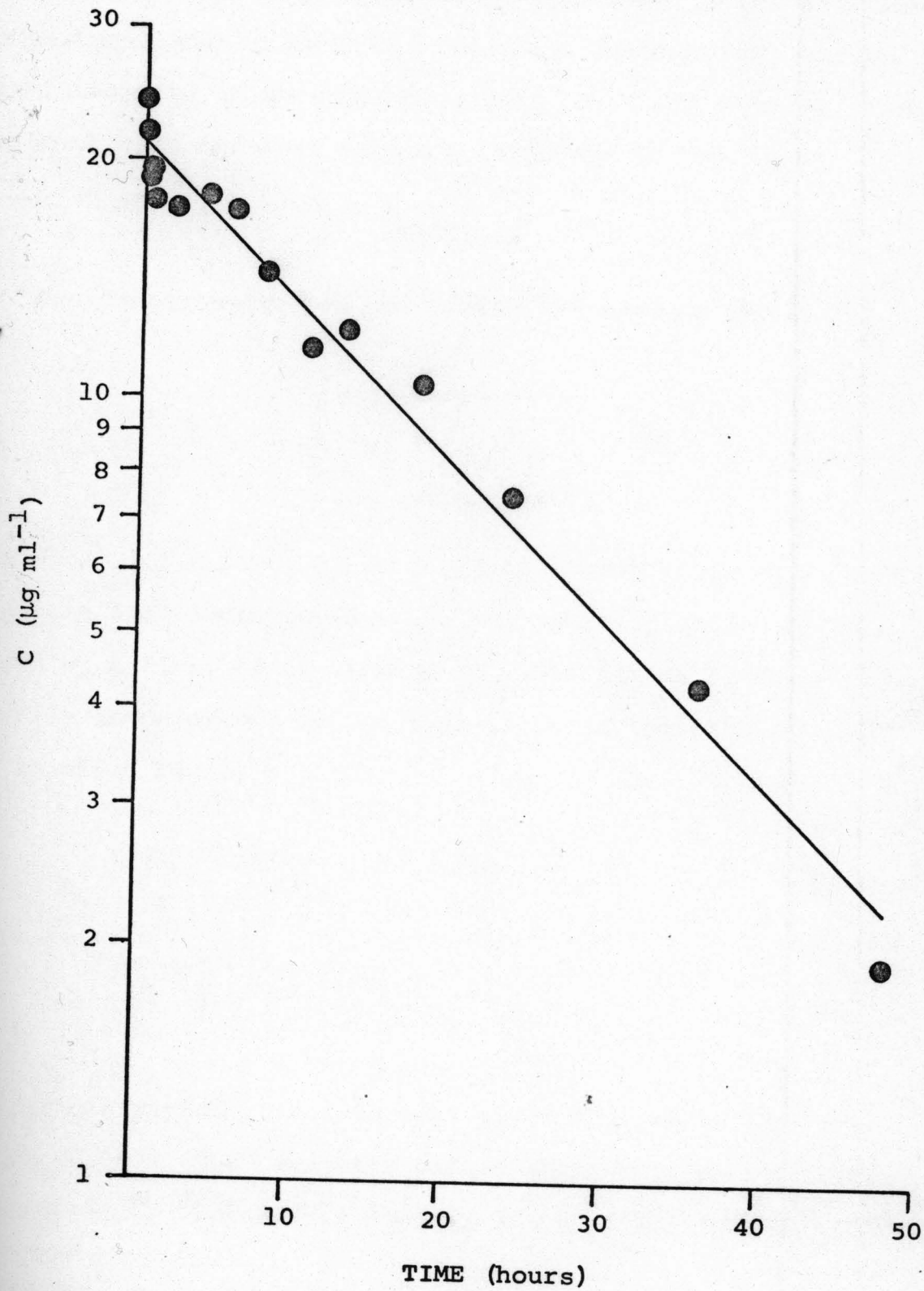


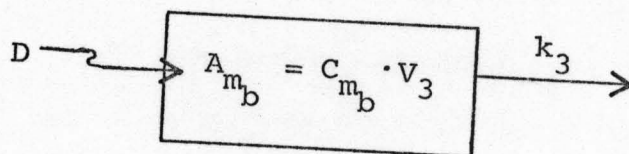
Figure 16. Semilogarithmic plot of metabolite concentration-time data of Table VII showing monoexponential decline in metabolite plasma level in dog B after dosing this compound. The linear regression equation including the 95% confidence limits on the slope and intercept is  $\log C = -0.0203 (\pm 0.0017)t + 1.3245 (\pm 0.0314)$ .



ascertain whether the metabolite obeyed one-, two-, or multi-compartment model kinetics. There was insufficient initial curvilinearity in the data to justify more than the one compartment open model as depicted in Scheme II (21).

Scheme II

One Compartment Open Model with Intravenous Injection



The dose  $D$  of metabolite is instantaneously injected into an apparent distribution volume  $V_3$  and excreted and/or metabolized by a first order process  $k_3$ . The plasma level vs time profile is described by equation 13 or in logarithmic form by equation 14 (21).

$$C_{m_b} = \frac{D}{V_3} e^{-k_3 t} \quad \text{Eq. 13}$$

$$\log C_{m_b} = \log \frac{D}{V_3} - \frac{k_3}{2.303} t \quad \text{Eq. 14}$$

The concentration axis intercept gives  $D/V_3$  while the slope is  $-k_3/2.303$ . The parameter values obtained from the linear regressions<sup>a</sup>, included in Figures 15-16, are summarized in

<sup>a</sup> Performed on a CompuCorp 344 Statistician Micro-Computer, Computer Design Corp., Los Angeles, California.

Table X and regression ANOVA data are summarized in Table XI.

As was expected, the biological half-life, equation 15, was long

$$t_{1/2} = \frac{0.693}{k_3}$$

Eq. 15

and was identical in the two experiments. The apparent distribution volumes were similar.

Since the appropriate pharmacokinetic models were thus established for both the methsuximide and metabolite data independently the drug and metabolite data from experiments A, B and C were analyzed as described below.

### C. Methsuximide Pharmacokinetics Including Formation of the Metabolite, 2-Methyl-2-phenylsuccinimide

#### 1. Development of the Model

Scheme III depicts the model representing intravenous administration of methsuximide with subsequent formation of the metabolite 2-methyl-2-phenylsuccinimide, where D is the amount of methsuximide intravenously administered,  $k_1$  and  $k_{-1}$  are as previously defined,  $k_2$  is the first order rate constant describing excretion and metabolism, other than to 2-methyl-2-phenylsuccinimide,  $k_f$  is the first order rate constant describing formation of the metabolite and  $k_3$  is the first order rate constant representing excretion and/or further metabolism of 2-methyl-2-phenylsuccinimide.

The differential equations 16-18 describe the rate of

Table X2-Methyl-2-phenylsuccinimide Pharmacokinetic Parameters

	<u>D/V<sub>3</sub> (mg/l)</u>	<u>V<sub>3</sub> (l)</u>	<u>k<sub>3</sub> (hr<sup>-1</sup>)</u>	<u>t<sub>1/2</sub> (hr)</u>
Experiment D	17.08	14.63	0.047	14.68
Experiment E	21.11	11.84	0.047	14.68

Table XIANOVA for Metabolite Linear RegressionsExperiment D

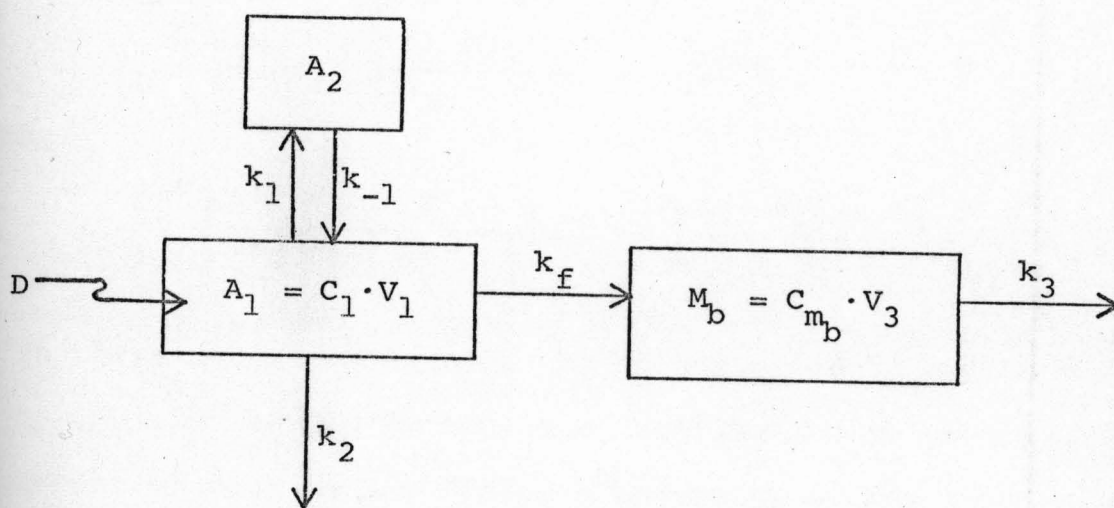
<u>Source</u>	<u>SS</u>	<u>df</u>	<u>MS</u>
regression	1.2993	1	1.2993
residual	0.0244	15	0.0016
total	1.3237	16	

Experiment E

<u>Source</u>	<u>SS</u>	<u>df</u>	<u>MS</u>
regression	1.2121	1	1.2121
residual	0.0247	13	0.0019
total	1.2368	14	

Scheme III

Model for a Drug Which Obeys Two Compartment Kinetics  
With a Metabolite Obeying One Compartment Kinetics



change of drug amounts in the various compartments as a function of time.

$$\frac{dA_1}{dt} = k_{-1}A_2 - (k_1 + k_f + k_2)A_1 \quad \text{Eq. 16}$$

$$\frac{dA_2}{dt} = k_1A_1 - k_{-1}A_2 \quad \text{Eq. 17}$$

$$\frac{dM_b}{dt} = k_fA_1 - k_3M_b \quad \text{Eq. 18}$$

The differential equations were integrated using Laplace transformations and matrix algebra, as described in Appendix B, to yield equations 19-21.

$$A_1 = \frac{D}{(\alpha - \beta)} \left\{ (k_{-1} - \beta)e^{-\beta t} - (k_{-1} - \alpha)e^{-\alpha t} \right\} \quad \text{Eq. 19}$$

$$A_2 = \frac{Dk_1}{(\alpha - \beta)} (e^{-\beta t} - e^{-\alpha t}) \quad \text{Eq. 20}$$

$$M_b = \frac{189}{203} Dk_f \left\{ \left[ \frac{k_{-1} - \alpha}{(k_3 - \alpha)(\beta - \alpha)} \right] e^{-\alpha t} + \right. \quad \text{Eq. 21}$$

$$\left. \left[ \frac{k_{-1} - \beta}{(k_3 - \beta)(\alpha - \beta)} \right] e^{-\beta t} + \left[ \frac{k_{-1} - k_3}{(\beta - k_3)(\alpha - k_3)} \right] e^{-k_3 t} \right\}$$

In these equations amounts can be converted to concentrations by dividing by the appropriate distribution volumes. The amount of drug in the tissue compartment at any time (equation 20) can only be calculated in this study since drug concentrations were determined only from the central compartment. Since the D in equation 21 is the dose of methsuximide then the factor 189/203, which is the ratio of molecular weights of metabolite:methsuximide, was necessary to convert  $\mu\text{g}$  of methsuximide to  $\mu\text{g}$  of metabolite. The composite rate constants  $\alpha$  and  $\beta$  are defined by equation 22 which is analogous to equation 6.

$$\frac{\alpha}{\beta} = \frac{1}{2} \left\{ (k_1 + k_f + k_2 + k_{-1}) \pm \sqrt{(k_1 + k_f + k_2 + k_{-1})^2 - 4(k_{-1}k_f + k_2k_{-1})} \right\} \quad \text{Eq. 22}$$

Equation 19 is identical to equation 5 with the  $k_2$  of Scheme I explicitly separated into  $k_2 + k_f$  in Scheme III. The

equation describing the concentration of metabolite in the body, equation 21, is triexponential indicating its dependence on the methsuximide biexponential process, equation 5, and its own monoexponential process, equation 13.

The stripping of triexponential curves is known to be an inaccurate procedure (22). In the case of the metabolite plasma levels vs time profiles, experiments A, B and C, however, initial estimates of some parameter values were provided by the final computer estimates from the methsuximide analysis, Table IX. Initial estimates of the rate constants  $\alpha$ ,  $\beta$  and  $k_{-1}$  were obtained in this fashion. There are two ways to obtain estimates of  $k_3$ . First,  $k_3$  can be estimated from the independent metabolite experiments or second, from the log-linear portion of equation 21. Since, in equation 21,  $\alpha > \beta > k_3$  there existed a time such that  $e^{-\alpha t}$  and  $e^{-\beta t}$  were insignificant relative to  $e^{-k_3 t}$  so that after this time equation 21 became log-linear with slope equal to  $-k_3/2.303$ . The volume  $V_3$  was estimated using the average  $V_3$ 's from experiments D and E. Initial estimates of  $k_f$  were obtained by comparison of areas under the metabolite and methsuximide plasma level profiles from each methsuximide experiment. The area under the  $C_1$  curve was obtained by integration of equation 19 as

$$\text{area } C_1 = \int_0^{\infty} C_1 dt \quad \text{Eq. 23}$$

resulting in equation 24 (20).

$$\text{area } C_1 = \frac{Dk_{-1}}{V_1\alpha\beta}$$

Eq. 24

The area under the metabolite curve was similarly given by equations 25 and 26 (see Appendix C).

$$\text{area } C_{m_b} = \int_0^{\infty} C_{m_b} dt$$

Eq. 25

$$= \frac{189k_f k_{-1} D}{203V_3 k_3 \alpha \beta}$$

Eq. 26

so that

$$k_f = \frac{203V_3 k_3}{189V_1} \cdot \frac{\text{area } C_{m_b}}{\text{area } C_1}$$

Eq. 27

These areas are from  $t_0$  to  $t_{\infty}$  so that the area under the curve from the time of the final data point  $C_f$  to infinity had to be calculated. If the  $C_f$ 's are at long enough times,  $t_f$ , such that equations 19 or 21 have become essentially monoexponential, then the concentrations after time  $t_f$  are given by:

$$C_1 = C_f' e^{-\beta t}$$

and

$$C_{m_b} = C_f'' e^{-k_3 t}$$

Eq. 28

where  $C_f'$  and  $C_f''$  represent the terminal data points for unchanged drug and metabolite, respectively. Integration from  $t_f$  to  $t_\infty$  then yields:

$$\text{terminal methsuximide areas} = \frac{C_f'}{\beta}$$

$$\text{Terminal metabolite areas} = \frac{C_f''}{k_3} \quad \text{Eq. 29}$$

The areas from time zero to  $t_f$  were determined by the trapezoidal rule. The remaining parameter  $k_2$  was estimated using equation 30

$$k_2 = \frac{\alpha\beta - k_{-1}k_f}{k_{-1}} \quad \text{Eq. 30}$$

which is obtained from the product of  $\alpha$  and  $\beta$  (equation 22).

The methsuximide and metabolite data, experiments A, B and C, were simultaneously fitted, using the program NREG, to equations 19 and 21. Initial estimates of parameters are given in Table XII.

## 2. NREG Computer Results

When all seven parameter values (Table XII) were allowed to float in the program wide confidence intervals were obtained for  $k_2$ ,  $k_f$  and  $D/V_3$ . High correlations ( $r > 0.80$ ) existed between these parameters. Since the independent metabolite experiments (Table XI) and curve stripping of metabolite data from experiments B and C yielded nearly

Table XII

## Initial Estimates of Scheme III Parameters

Parameter <sup>a</sup>	Experiment A	Experiment B	Experiment C	Source
$k_1$	2.80	3.67	1.86	Table IX
$k_{-1}$	1.47	3.40	3.28	Table IX
$k_2$	0.38	0.96	0.55	Eq. 30
$k_f$	0.26	0.72	0.29	Eq. 27
$k_3$	0.047	0.047	0.047	Table XI <sup>b</sup>
$D/V_1$	23.3	18.5	11.1	Table IX
$D/V_3$	18.9	18.9	18.9	Table XI

<sup>a</sup>Units of all rate constants =  $\text{hr}^{-1}$ ; units of  $D/V = \mu\text{g ml}^{-1}$ .

<sup>b</sup>For experiments B and C the terminal log linear form of eq. 21 yielded  $k_3$  values of 0.047 and 0.050  $\text{hr}^{-1}$  respectively.

identical values of  $k_3$ , and since the value of  $D/V_3$  was accurately determined from experiments D and E these parameters were fixed during the computer analysis. This procedure significantly reduced the 95% confidence intervals while causing only slight changes in the values of the remaining parameters.

The computer was unable to satisfactorily fit the metabolite data from data set A regardless of whether parameters were fixed or not. The primary reason for this failure was that this experiment was terminated at 12 hours. The values of  $\alpha$ ,  $\beta$  and  $k_3$  were such that equation 21 never became monoexponential. The rate constant  $k_3$  could not therefore be computer determined and  $k_f$ , calculated with equation 27, was in error because equation 29 was not applicable. Another reason for this failure is that the initial estimate of  $D/V_3$  was based on experiments in Dog B whereas data set A was from Dog A. Since the Table IX parameters show variability between data sets the assumption that  $V_3$  was the same in both dogs was tenuous.

The final computer estimates of experiment B and C parameters are listed in Table XIII with relevant statistics and derived pharmacokinetic parameters. The solid lines in Figures 7-9 are the computer fits to the plasma level vs time equations as specified with the exception of the Figure 7 metabolite profile which is hand drawn. All data was weighted by the factor  $1/C$ .

It is noteworthy that the incorporation of the

Summary of Pharmacokinetic Computer Estimated Scheme III Parameters

Parameter	Data Set B	Data Set C
$k_1$ ( $\text{hr}^{-1}$ )	3.84 (0.52-7.17) <sup>a</sup>	0.71 (0.20-1.23)
$k_{-1}$ ( $\text{hr}^{-1}$ )	3.80 (1.92-5.68)	1.14 (0.26-2.02)
$k_2$ ( $\text{hr}^{-1}$ )	1.01 (0.69-1.33)	0.50 (0.42-0.58)
$k_f$ ( $\text{hr}^{-1}$ )	0.77 (0.54-1.00)	0.28 (0.24-0.31)
$k_3$ (F) <sup>b</sup> ( $\text{hr}^{-1}$ )	0.047	0.047
$D/V_1$ ( $\mu\text{g ml}^{-1}$ )	18.9 (12.8-25.0)	10.4 (9.3-11.5)
$D/V_3$ (F) <sup>b</sup> ( $\mu\text{g ml}^{-1}$ )	18.9	18.9
$V_1$ (l)	13.2	24.0
$V_3$ (l)	13.2	13.2
$\alpha$ ( $\text{hr}^{-1}$ )	8.64	2.24
$\beta$ ( $\text{hr}^{-1}$ )	0.78	0.40
$t_{1/2}$ (methsuximide)	0.89	1.73
$t_{1/2}$ (metabolite)	14.74	14.74
Initial sums of squares	0.86	1.13
Final sums of squares	0.78	1.01

Table XIII (continued)

Parameter	Data Set B	Data Set C
R <sup>2</sup> (methsuximide)	0.9979	0.9937
r (methsuximide)	0.9951	0.9931
R <sup>2</sup> (metabolite)	0.9919	0.9914
r (metabolite)	0.9933	0.9928
R <sup>2</sup> (overall)	0.9950	0.9928

<sup>a</sup> 95% confidence interval.

<sup>b</sup> Indicates parameter was fixed.

metabolite equation in the computer program did not significantly alter the methsuximide parameters (cf. Table IX). Of particular interest is the relative importance of the rate of biotransformation of methsuximide to the pharmacologically active and potentially toxic 2-methyl-2-phenylsuccinimide,  $k_f$ , compared to the overall elimination and metabolism rate constants,  $k_f + k_2$ . This metabolic step accounted for 43 and 36% of the overall elimination of methsuximide in the two experiments. The large value of  $k_f$  together with the small  $k_3$  (long half-life) leads to a metabolite plasma level profile after a single intravenous dose of methsuximide which mirrors a prolonged sustained release type profile. Figures 7-9 indicate that the metabolite appeared in the dog's plasma in less than five minutes after intravenous methsuximide dosing. Since methsuximide is orally administered on a daily basis to petit mal patients there may be significant accumulation of the metabolite in the body with multiple dosing regimens.

Methsuximide administration results in induction of hepatic drug metabolizing enzymes (5, 6). It has been observed in patients on chronic methsuximide therapy that this 'auto-induction' process is so marked that 2-methyl-2-phenylsuccinimide is the only drug related compound detected in blood after repeated oral methsuximide administration, even shortly after dosing (23). The possibility exists that the metabolite may be partially or totally responsible for the enzyme induction. If this were the case, then both rate

constants,  $k_f$  and  $k_3$ , may increase after repeated exposure to methsuximide. If  $k_3$  were to increase then accumulation of metabolite in the body would be reduced. The above problem could be elucidated pharmacokinetically through multiple dosing experiments with methsuximide and 2-methyl-2-phenylsuccinimide. However, since several other metabolites have been found in dog plasma (11) a complete pharmacokinetic analysis would require administration of each metabolite individually. This is the approach for which further experiments should be designed.

## APPENDIX A

Example of Computer Output Using the Program NREG



```

00102 DIMENSION TH(1),Y(NRPD,1),T(1)
00103 COMMON T
00104 COMMON/LIM/NEXP1,NEXP2,K,AL,BE
00105 TH1 = TH(1)
00106 TH2 = TH(2)
00107 TH3 = TH(3)
00110 TH4 = TH(4)
00111 TH5 = TH(5)
00112 TH6 = TH(6)
00113 TH7 = TH(7)
00114 AL = 0.5*((TH1+TH4+TH3+TH2)+SQRT((TH1+TH4+TH3+TH2)**2-4*(TH2*TH4+
00115 TH3*TH2)))
00116 BE = 0.5*((TH1+TH4+TH3+TH2)-SQRT((TH1+TH4+TH3+TH2)**2-4*(TH2*TH4+
00117 TH3*TH2)))
00121 DO 10 I = 1,NEXP1
00122 10 Y(I,1) = (TH6/(AL-BE))*((TH2-BE)*EXP(-BE*T(I)))-(TH2-AL)*EXP(-AL*T
00123 I(I))
00125 IF( K.EQ.NEXP1) GO TO 30
00126 DO 20 I = K,NEXP
00130 20 Y(I,1) = 189./203.*TH7*TH4*((TH2-AL)/((TH5-AL)*(BE-AL)))*EXP(-AL*T
00131 I(I)) + ((TH2-BE)/(AL-BE))*((TH5-BE)*EXP(-BE*T(I)))*((TH2-TH5)/(BE-
00132 2*TH5))*(AL-TH5))*EXP(-TH5*T(I))
00133 30 CONTINUE
00134 RETURN
00135 END

```

```

END OF COMPILATION: NO DIAGNOSTICS.
FOR=SI
FORTRAN-MACC 1.145-07/30/75-13:27:58 OTP
00101 SUBROUTINE OUTPUT,NEXP,NEXP2,TH1,NVAR,NP,NRPD,YPRD,YOBS)
00102 DIMENSION T(1),YOBS(1),YPRD(1)
00103 COMMON Y
00104 COMMON/LIM/NEXP1,NEXP2,K,AL,BE
00105 K = 0
00106 200 CONTINUE
00107 S02 = 0.0
00110 SP2 = 0.0
00111 SC = 0.0
00112 SYORS = 0.0
00113 SOEV = 0.0
00114 COR = 0.0
00115 RRS = 0.0
00116 YOH = 0.0
00117 YPH = 0.0
00120 K = K + 1
00121 IF (K.EQ.1) GO TO 210
00122 LL = REXP1 + 1
00123 LU = NEXP
00124 LU = NEXP
00125 GO TO 250
00126 210 LL = 1
00127 LU = NEXP1
00130 250 DO 10 I = LL,LU
00133 SYOBS = YOBS + YOBS(I)**2
00134 SDEV = SDEV + (YOBS(I)-YPRD(I))**2
00135 YOH = YOH + YOBS(I)
00136 YPH = YPH + YPRD(I)
00137 JO CONTINUE

```

```

00141 29. RS = (SYOBS-SDEV)/SYOBS
00142 30. YOM=YOM/NEXP
00143 31. YPM=YPM/NEXP
00144 32. DO 20 I=1,NEXP
00150 33. SC = SC+(YOBS(I)-YOM)*(YPRD(I)-YPM)
00151 34. SO2 = SO2 + (YOBS(I)-YOM)**2
00152 35. SP2 = SP2 + (YPRD(I)-YPM)**2
00153 36.
00154 37. 20 CONTINUE
00155 38. COR = SC/(SQRT(SO2))*SQRT(SP2)
00163 39. WRITE(6,100) K,RS,K,COR
00164 40. 100 FORMAT(1H0,'R-SQUARED FOR FUNCTION',11,' =',F6.4)
00170 41. JION=11,'F,6.4)
00171 42. IF (INXP2.EQ.0) GO TO 66
00172 43. IF (K.GT.1) GO TO 300
00173 44. SYOBS1 = SYOBS
00174 45. SDEU1 = SDEV
00175 46. GO TO 200
00176 47. 300 SYOBS = SYOBS + SYOBS1
00177 48. SDEV = SDEV + SDEU1
00178 49. RS = (SYOBS - SDEV)/SYOBS
00179 50. WRITE(6,111) RS
00180 51. 111 FORMAT(1X,'OVER ALL R-SQUARED=',F6.4)
00201 52. 66 CONTINUE
00202 53. WRITE (6,101) ALABE
00210 54. 101 FORMAT(1X,'ALPHA =',F10.5)
00211 55. CALL PRPL (NEXP,NEXPD,THFN,NVAR,NP,NRPRD,YPRD,YOBS)
00212 56. RETURN
00213 57. END

```

```

END OF COMPILATION: PRPL NO DIAGNOSTICS.
FOR:SI
FORTRAN=MACC 1195-07/30/75-13:28:09 PRPL
00101 1. SUBROUTINE PRPL (NEXP,NEXPD,THFN,NVAR,NP,NRPRD,YPRD,YOBS)
00102 2. DIMENSION T(1),YOBS(NRPRD,1),YPRD(NRPRD,1)
00103 3. COMMON /
00104 4. COMMON/LIM/NEXPI,NEXP2,K
00105 5. MM = 0
00106 6. 200 CONTINUE
00107 7. MM = MM + 1
00110 8. IF (MM.EQ.1) GO TO 50
00112 9. DO 140 I = 1,NEXP2
00115 10. MM = NEXPI + 1
00116 11. YOBS(I,1) = YOBS(MM,1)
00117 12. 140 T(I) = T(MM)
00121 13. LU = NEXP2
00122 14. GO TO 60
00123 15. 50 CONTINUE
00124 16. LU = NEXPI
00125 17. 60 CONTINUE
00126 18. YMAX = YOBS(1,1)
00127 19. DO 10 I = 2,LU
00132 20. 10 YMAX = AMAX1(YMAX,YOBS(I,1))
00134 21. IF (MM.EQ.2) GO TO 170
00136 22. THAX = T(I)
00137 23. DO 15 I = 2,NEXP
00142 24. 15 THAX = AMAX1(THAX,T(I))
00144 25. BRAX = THAX/(NEXP2-NEXP2)

```

```

00145 26. NM1 = NEXPI + NEXP2 + 1
00146 27. NM2 = NEXPI + NEXP2 + 2
00147 28. TUNMI) = 0.0
00150 29. DO 30 I = NM2,NEXP2
00153 30. J = I - 1
00154 31. 30 T(I) = T(J) + BMAX
00156 32. IF ( MM.EQ.1 ) GO TO 160
00160 33. GO TO 170
00161 34. 160. NEXI = NEXPI
00162 35. NEXPI = NEXP2
00163 36. K = NEXP2
00164 37. GO TO 180
00165 38. 170 CONTINUE
00166 39. NEXPI = I
00167 40. K = I
00170 41. NEXP = NEXP2
00171 42. 180 CONTINUE
00172 43. CALL F(THIN,YPRD,NEXPD,NVAR,NP,NRPRD)
00173 44. NEXPI = NEXI
00174 45. DO 20 I = 1,NEXP2
00177 46. 20 YMAX = AMAXI(YMAX,YPRD(I),I)
00201 47. YSCALE = YMAX/8.8
00202 48. TSCALE = TMAX/8.8
00203 49. CALL GRPH2MT,IHR,YPRD,IHR,NEXP2,3HXS,0,YSCL,0,YSCL,17HFUNCTIO
00203 50. IN VALUES...6MTIME...198 PLOT OF OBSERVED,0'S, AND PREDICTED,0'S, V
00204 51. 2ALUES,1H*)
00205 52. CALL GRPH2V(I,IHR,YOBS,IHR,'LULHSAME,1HQ)
00206 53. CALL GRPHND
00206 54. IF (NEXP2.EQ.0) GO TO 66
00210 55. IF (MM.EQ.1) GO TO 200
00212 56. 66 CONTINUE
00213 57. RETURN
00214 58. END
    
```

END OF COMPILATION: NO DIAGNOSTICS.

MAP 017P-07730-13128

ADDRESS LIMITS 001000 047633 050000 072543  
 STARTING ADDRESS 047411  
 WORDS DECIMAL 19868 IBANK 9572 DBANK

SEGMENT	SWAINS	001000	047633	050000	072543
NEXP13/NAGFORFUNIO	I	001000	001047	0	050000 050000
UR3SRH/UWCC	I	001050	001126		
UR2MSK/UWCC	I	001127	001260	0	050001 050024

URTRI/UKCCO	1	001261	001446	2	BLANK&COMMON
URSORT/NAGUTLITV02	1	001447	002266	0	050027 050166
NRIFK/NAGNLINREG00	1	002267	002530	0	050167 050247
NRIRD/NAGNLINREG00	1	002531	002632	2	BLANK&COMMON
NRIRT/NAGNLINREG00	1	002633	002753	2	BLANK&COMMON
NRIDR/NAGNLINREG00	1	002754	003033	2	BLANK&COMMON
ASTRCSS/NAGFORFUNIT0	1	003034	003257	0	050402 050417
NRILT/NAGNLINREG01	1	003260	003430	0	050420 050445
NRISWP/NAGNLINREG00	3	NRIBLK		0	050446 050477
NRISJ/NAGNLINREG00	1	003431	004054	2	BLANK&COMMON
NRISZ/NAGNLINREG00	1	004055	004343	2	BLANK&COMMON
NRITV/NAGNLINREG00	1	004344	004742	0	050546 050616
NRISRT/NAGNLINREG00	1	004743	005172	2	BLANK&COMMON
NRISVD/NAGNLINREG00	1	005173	007312	2	BLANK&COMMON
NRITRS/NAGNLINREG00	1	007313	010147	2	BLANK&COMMON
NRIDIF/NAGNLINREG04	1	010150	010744	2	BLANK&COMMON
NRICK/NAGNLINREG00	1	010745	010762	0	051347 051443
URPHID/UMCC	1	010763	010771	0	051444 051444
URDATE/NAGUTILITY00	1	010772	011010	0	051445 051445
URTIME/NAGUTILITY00	1	011011	011017	0	051446 051447
EOUTB/SYS	1	011020	012056	0	051450 051450
NR1HO/NAGNLINREG04	1	012057	012622	2	051451 051451
NR1HC/NAGNLINREG06	1	012623	015362	0	052035 052557
NR1NST/NAGNLINREG01	1	015363	017065	0	052540 053447
NR1FPV/NAGNLINREG00	3	NRIBLK		0	053450 054264
NR1BLK (COMMON BLOCK)	1	017066	017273	2	BLANK&COMMON
NR1WK/NAGNLINREG02	1	017274	021737	0	054265 054372
NR1HV/NAGNLINREG02	3	NRIBLK		2	BLANK&COMMON
NR1PUT/NAGNLINREG00	1	021740	022703	0	054411 054657
NR1BDS/NAGNLINREG01	1	022704	023037	0	BLANK&COMMON
NR1BDM/NAGNLINREG00	1	023040	023123	2	055036 055061
NR1LTN/NAGNLINREG00	1	023124	023201	0	BLANK&COMMON
NR1BDA/NAGNLINREG00	1	023202	023277	2	055062 055113
NR1LTZ/NAGNLINREG00	1	023300	023351	0	055114 055127
NR1GRD/NAGNLINREG01	1	023352	023436	2	BLANK&COMMON
NR1PST/NAGNLINREG00	1	023437	023577	0	055207 055231
				2	BLANK&COMMON
				0	055232 055265
				2	BLANK&COMMON
				0	055266 055422
				2	BLANK&COMMON

HRTMS/HAGLHREGO1	1	02425	024311	0	055423	055436
NISYMS/FORIO	2			2	BLANK\$COMMON	
NEP5\$/HAGFORFUN11	1	024312	024407	2	055437	055443
ALGS\$/HAGFORFUN10	1	024410	024517	2	055444	055453
GRPRT/GRFPRTUW01	1	024520	024635	2	055454	055527
	3	024636	025123	0	055530	055706
GRMCH/GRFPRTUW01	1	025124	025133	2	BLANK\$COMMON	
URSRCH/UWCC01	1	025134	025426	2	055707	055710
NFTCH\$/FORIO	1	025427	025730	2	055711	055716
NRIED\$/HAGLHREGO9	1	025731	031470	2	055717	055717
URXIT\$/HAGUTILITV01	1	031471	031550	0	055720	055755
ERUS	1			2	055756	057345
NOTUS\$/FORIO	1	031551	032060	2	057346	057404
FOI052\$/FORIO	1	032061	034437	2	057405	057411
NTABS\$/FOPIO	1			2	057412	062076
NEFTI\$/FORIO	1	034440	035514	2	062077	062143
NEFT0\$/FORIO	1	035515	036105	2	062144	062374
NO5Y\$/FCRIO	1	036106	036215	2	062375	062436
UMERR\$/HAGFORFUN02	1	036216	037026	2	062437	062442
CNTRHD/GRFPRTUW01	1	037027	037256	2	062443	063102
	2			0	063103	063143
GRAPHZ/HAGGRAPH202	1	037257	041435	2	BLANK\$COMMON	
URNPAR/UWCC	1	041436	041464	2	063144	063516
FOI054\$/FORIO	1	041465	041764	2	063517	063517
HREG/HAGLHREGO2	1	041765	042110	2	063520	063521
EXP\$/HAGFORFUN10	1	042111	042213	2	063522	063543
NOUP\$/FORIO	1	042214	042271	2	063544	063556
SQPT\$/HAGFORFUN10	1	042272	042341	2	063557	063602
FOI051\$/FORIO	1	042342	045534	2	063603	063603
G17345 (COMMON BLOCK)	1	045535	046265	2	063604	063615
GRAPHZ/GRFPRTUW02	1			2	063616	064715
	3	046266	046635	0	064716	070056
LIM (COMMON BLOCK)	1	070057	070271	2	BLANK\$COMMON	
BLANK\$COMMON (COMMON BLOCK)	1	070272	070276	2	070272	070276
PRPL	1	070277	070360	2	070277	070360
OTP	1	LIM		0	070361	070462
	3	046636	047142	2	BLANK\$COMMON	
SURF	1	LIM		0	070463	070561
	3	047143	047410	2	BLANK\$COMMON	
MAIN	1	LIM		0	070562	070633
	3	047411	047633	2	BLANK\$COMMON	
	3	LIM		0	070634	072543
	3			2	BLANK\$COMMON	

SYSELIBS, LEVEL 19  
 END OF COLLECTION - TIME 4.155 SECONDS

6X9T

SUMMARY OF THE DATA

ITEMS

ITEMS(1): NEXP = 23 NUMBER OF EXPERIMENTS  
 ITEMS(2): NVAR = 1 NUMBER OF VARIABLES  
 ITEMS(3): NP = 7 NUMBER OF PARAMETERS  
 ITEMS(4): NRORS = 23 ROW DIMENSION OF ARRAY YORS  
 ITEMS(5): NRPRD = 23 ROW DIMENSION OF ARRAY YPRD  
 ITEMS(6): NRWTS = 23 ROW DIMENSION OF ARRAY WTS  
 ITEMS(7): METHOD = 'MARQ' MARQUARDT'S METHOD  
 ITEMS(8): KWTS = 'ALL' WEIGHTING BY OBSERVATION  
 ITEMS(9): KDERIV = 'NREG' NREG CALCULATED DERIVATIVES  
 ITEMS(10): KSYM = 'CENT' CENTRAL DIFFERENCES  
 ITEMS(11): KDIFP = 'REL' RELATIVE DIFFERENCES  
 ITEMS(12): MAXIT = 50 LIMIT ON NUMBER OF ITERATIONS

TOLERANCES

TOL(1) = 1.500000-06 REL. CHANGE IN A PARAMETER  
 TOL(2) = 1.000000-09 REL. CHANGE IN SUM OF SQUARES  
 TOL(3) = 0.000000 RATIO TO INITIAL SUM OF SQUARES  
 TOL(4) = 1.000000-16 PIVOT TOLERANCE

SUNDRIES

NUMBER OF ACTIVE PARAMETERS = 5  
 NUMBER OF CELLS OF SCRATCH REQUIRED = 471

PARAMETERS

NO.	NAME	INITIAL VALUE	PROPORTION FOR DERIV. ESTIMATE	LOWER BOUND	UPPER BOUND
1	K1	2.8000000+00	1.0000000-03	1.0000000-04	1.0000000+04
2	K-1	1.4700000+00	1.0000000-03	1.0000000-04	1.0000000+04
3	K2	3.8000000-01	1.0000000-03	1.0000000-04	1.0000000+04
4	KF	2.4000000-01	1.0000000-03	1.0000000-04	1.0000000+04
5	K3	4.7000000-02	1.0000000-03	1.0000000-04	1.0000000+04
6	DV1	2.3300000+01	1.0000000-03	1.0000000-04	1.0000000+04
7	DV3	1.8900000+01	1.0000000-03	1.0000000-04	1.0000000+04



THE ITERATIONS

ITERATION NO. 1	BASE POINT	TEST POINT
SUM OF SQUARES	2.0451071+01	1.2642243+01
L	0	0
LAMBDA	0.0000000	0.0000000
GAMMA	1.0000000+00	1.0000000+00
ANGLE IN DEGREES	.0000	88.8377
MAX. PIVOT REDUCTION	1.0000000+00	1.4509598-01
PAR.		
1 K1	2.8000000+00	4.1919542+00
2 K-1	1.4700000+00	1.5587288+00
3 K2	3.3000200-01	4.6122164-01
4 KF	2.6000000-01	5.1864593-01
5 K3	F 4.7000000-02	F 4.7000000-02
6 DV1	2.3300000+01	3.2726709+01
7 DV3	F 1.8900000+01	F 1.8900000+01

CUMULATIVE NO. OF FUNCTION CALLS = 2 ITERATION TIME = .170 SECONDS CUMULATIVE TIME = .339 SECONDS

ITERATION NO. 10	BASE POINT	TEST POINT
SUM OF SQUARES	3.3377034+00	3.3376852+00
L	0	0
LAMBDA	0.0000000	0.0000000
GAMMA	1.0000000+00	1.0000000+00
ANGLE IN DEGREES	84.5875	89.9846
MAX. PIVOT REDUCTION	1.8181388-01	1.0447983-01
PAR.		
1 K1	1.6108884+01	1.6103229+01
2 K-1	S 4.9103313-01	4.9026747-01
3 K2	8.3745060+00	8.3835536+00
4 KF	8.2606049+00	S 8.2662183+00
5 K3	F 4.7000000-02	F 4.7000000-02
6 DV1	1.1337907+03	1.1361034+03
7 DV3	F 1.8900000+01	F 1.8900000+01

CUMULATIVE NO. OF FUNCTION CALLS = 14 ITERATION TIME = .071 SECONDS CUMULATIVE TIME = 1.006 SECONDS

THE LAST ITERATION

ITERATION NO. 13      BASE POINT      TEST POINT  
 SUM OF SQUARES    3.3376848+00      3.3376848+00  
 LAMBDA            0                    18  
 GAMMA            0.0000000            7.6169012+01  
 ANGLE IN DEGREES 1.0000000+00      5.0000000-01  
 MAX. PIVOT REDUCTION 89.9838            11.8430  
                          6.2900740-02      1.6006252+00

PAR.    1 K1            1.6102946+01  
          2 K-1          4.9013491-01    S  
          3 K2            8.3851470+00  
          4 KF            8.2674358+00  
          5 K3            4.7000000-02    F  
          6 DV1          1.1365464+03  
          7 DV3            1.8900000+01    F

CUMULATIVE NO. OF FUNCTION CALLS = 36      ITERATION TIME = .366 SECONDS      CUMULATIVE TIME = 1.658 SECONDS

ITERATION TERMINATES:  
 MAX. RELATIVE CHANGE IN A PARAMETER .LT. TOL(1) = 1.5000000-06

THE LAST ITERATION

WADSON ACADEMIC COMPUTING CENTER - SUBROUTINE NREG METHSUXIMIDE AND METABOLITE DATA 1-24-74 WEIGHTED BY 1/C 07/30/75 TRANSMITTAL NO. B15365 CALL 1 PAGE 4

MADISON ACADEMIC COMPUTING CENTER - SUBROUTINE NREG  
 METHSOXIMIDE AND METABOLITE DATA 1-24-74 WEIGHTED BY 1/C

07/30/75 TRANSMITTAL NO. B19365 CALL 1 PAGE 5

FINAL PARAMETER VALUES

SUM OF SQUARES = 3.3376848+00

AR.	PARAMETER	FINAL VALUE
1	K1	1.6102946+01
2	K-1	4.9013491-01
3	K2	8.3851470+00
4	KF	8.2674358+00
5	K3	4.7000000-02
6	DV1	1.1365464+03
7	DV3	1.68900000+01

ITERATION SUMMARY

ITER NO.	SUM OF SQUARES	L	LAMRDA	GAMMA	ANGLE	NO. PIVOT FAILURES	NO. BOUNDS FAILURES	MAX. PIVOT REDUCTION	TIME IN SECONDS
0	2.0451071+01	0	0.0000000	1.0000000+00	.0000	0	0	1.0000000+00	.000
1	1.2642243+01	0	0.0000000	1.0000000+00	88.8377	0	0	1.4509598-01	.170
2	1.0717873+01	0	0.0000000	1.0000000+00	89.4822	0	0	3.8250508-02	.144
3	1.0602297+01	0	0.0000000	1.0000000+00	89.6044	0	0	2.4996527-02	.072
4	7.8271885+00	2	1.3070227-04	1.0000000+00	89.4272	0	0	2.1107513-01	.103
5	5.5815112+00	1	4.3552835-05	1.0000000+00	89.8060	0	0	2.6315963-01	.071
6	4.7656243+00	1	1.1200597-05	1.0000000+00	89.8844	0	0	2.3161651-01	.086
7	4.6951181+00	0	0.0000000	1.0000000+00	89.9823	0	0	1.7335901-01	.070
8	3.3492301+00	0	0.0000000	1.0000000+00	89.9859	0	0	1.1476095-01	.069
9	3.3377034+00	0	0.0000000	1.0000000+00	84.5895	0	0	1.8181388-01	.070
10	3.3376852+00	0	0.0000000	1.0000000+00	89.9446	0	0	1.0447983-01	.071
11	3.3376849+00	0	0.0000000	1.0000000+00	89.9827	0	0	6.0803708-02	.128
12	3.3376848+00	0	0.0000000	1.0000000+00	89.9838	0	0	6.2900740-02	.067
13	3.3376848+00	18	7.6169012+01	5.0000000-01	11.8430	0	0	1.6006252+00	.366

MANUSCRIPT ACCEPTED FOR PUBLICATION BY THE JOURNAL OF PHARMACEUTICAL SCIENCES

SINGULAR VALUES AND VECTORS AT TERMINAL POINT

SING. VALUES	1.1274202+01	8.3881985-01	5.6328676+01	2.7297219-01	9.8074986-04
PAR. 1 K1	.0116743	-.1127463	.1265469	-.9854616	.0017625
2 K-1	-.9988489	.0269920	.0383708	-.0099934	.0001672
3 K2	.0397844	.1125778	.9862397	.1142290	.00051276
4 KF	.0241137	.9868486	.0991181	-.1253532	-.0032390
F 5 K3	...	...	...	...	...
6 DVI	-.1127463	.1265469	-.9854616	.0017625	-.27836.8535156
F 7 DV3	...	...	...	...	...

NORMALIZING ELEMENTS AND CORRELATION MATRIX

PARAMETER NO.	1	2	3	4	5
NORM. ELTS.	4.0411890+00	2.0960960-01	5.5311241+00	3.5402238+00	1.0196076+03

PAR.	1 K1	2 K-1	3 K2	4 KF	5 K3	6 DVI	7 DV3
1 K1	1.0000000						
2 K-1	.5306135	1.0000000					
3 K2	-.4711510	-.6757723	1.0000000				
4 KF	-.3127974	-.7015691	.8643170	1.0000000			
F 5 K3	...	...	...	...	1.0000000		
6 DVI	-.4447006	-.8134292	.9452399	-.9328822	...	1.0000000	
F 7 DV3	...	...	...	...	...	...	1.0000000

PARAMETER NO. 1 2 3 4 5 6 7

NORM. ELTS. ...

PAR. F 7 DV3 ...

LIMIT VALUE = 3.32398E+00

LIMIT VALUE = 3.32398E+00

PARAMETER NO. 1 2 3 4 5 6 7

CONFIDENCE LIMITS ON LINEAR HYPOTHESIS

PAR.	K1	K2	K3	DV1	DV3	LOWER LIMIT	FINAL VALUE	UPPER LIMIT
1	K1					1.262275+01	1.6102946+01	1.9583317+01
2	K-1					3.0961398-01	4.9703491-01	6.7065585-01
3	K2					3.6216072+00	8.3851470+00	1.3148687+01
4	KF					5.2185082+00	8.2674358+00	1.1316363+01
5	K3					..	..	..
6	DV1					2.5843527+02	1.1365464+03	2.0146575+03
7	DV3					..	..	..
						1.8900000+01		

E-X-P-L-O-R-A-T-I-O-N

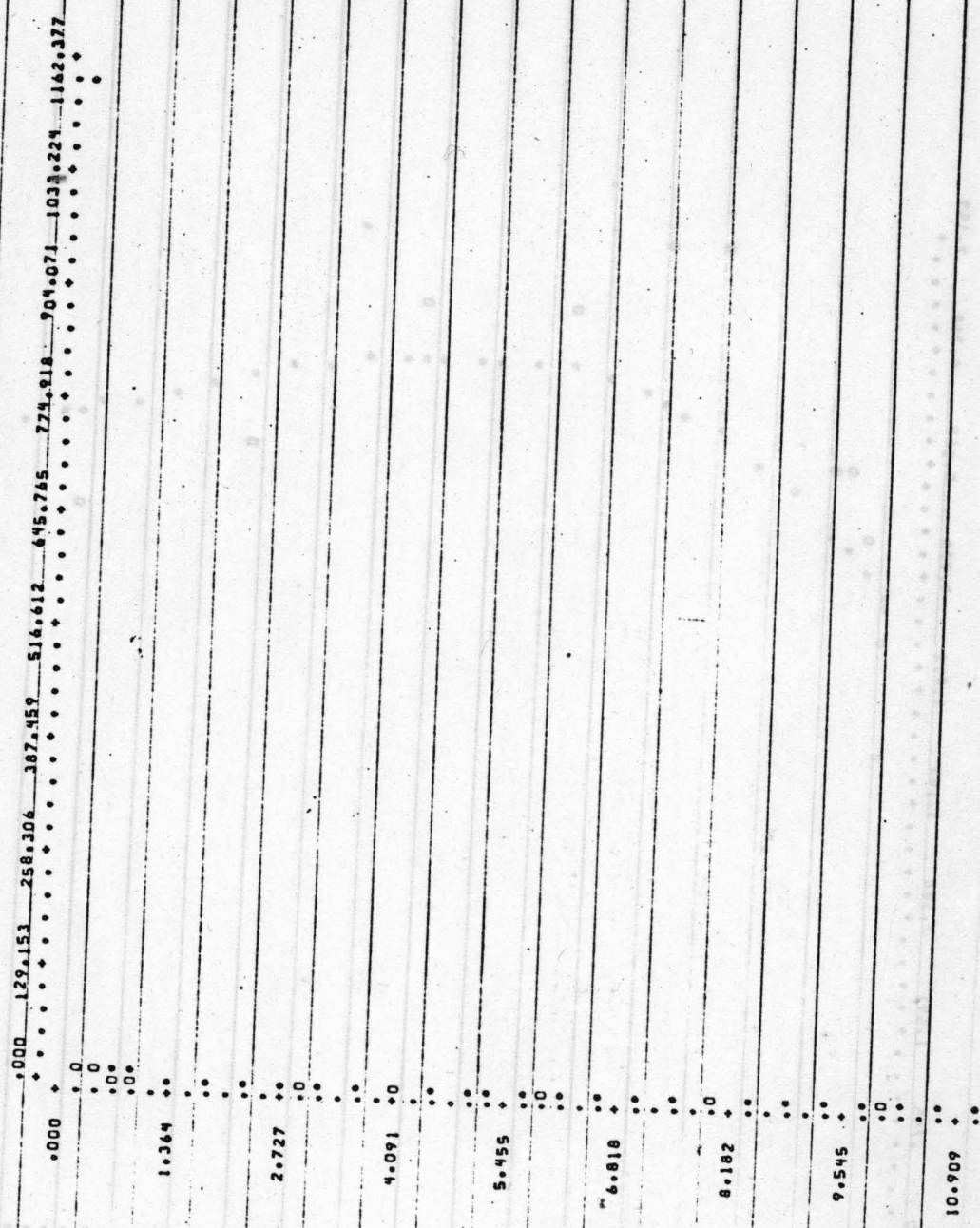
PAR.	K1	K2	K3	DV1	DV3	LOWER TEST	SUM OF SQUARES	LINEAR EST. OF SUM OF SQUARES	UPPER TEST	SUM OF SQUARES	LINEAR EST. OF SUM OF SQUARES
1	K1					1.5232853+01	3.4220665+00	3.4162008+00	1.6973038+01	3.4074404+00	3.4162008+00
2	K-1					4.4500468-01	3.5944698+00	3.5959760+00	5.3526514-01	3.5777870+00	3.5959760+00
3	K2					7.1942820+00	4.1854282+00	4.0747271+00	9.5760319+00	3.9781567+00	4.0747271+00
4	KF					7.5052040+00	3.8148378+00	3.7812358+00	9.0296676+00	3.7438282+00	3.7812358+00
5	K3					4.7000000-02	..	..	4.7000000-02	..	..
6	DV1					9.1701842+02	5.3668222+00	5.3667911+00	1.3560742+03	5.3667962+00	5.3667911+00
7	DV3					1.8900000+01	..	..	1.8900000+01	..	..

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FUNCTION VALUES

PLOT OF OBSERVED, O'S, AND PREDICTED, P'S, VALUES

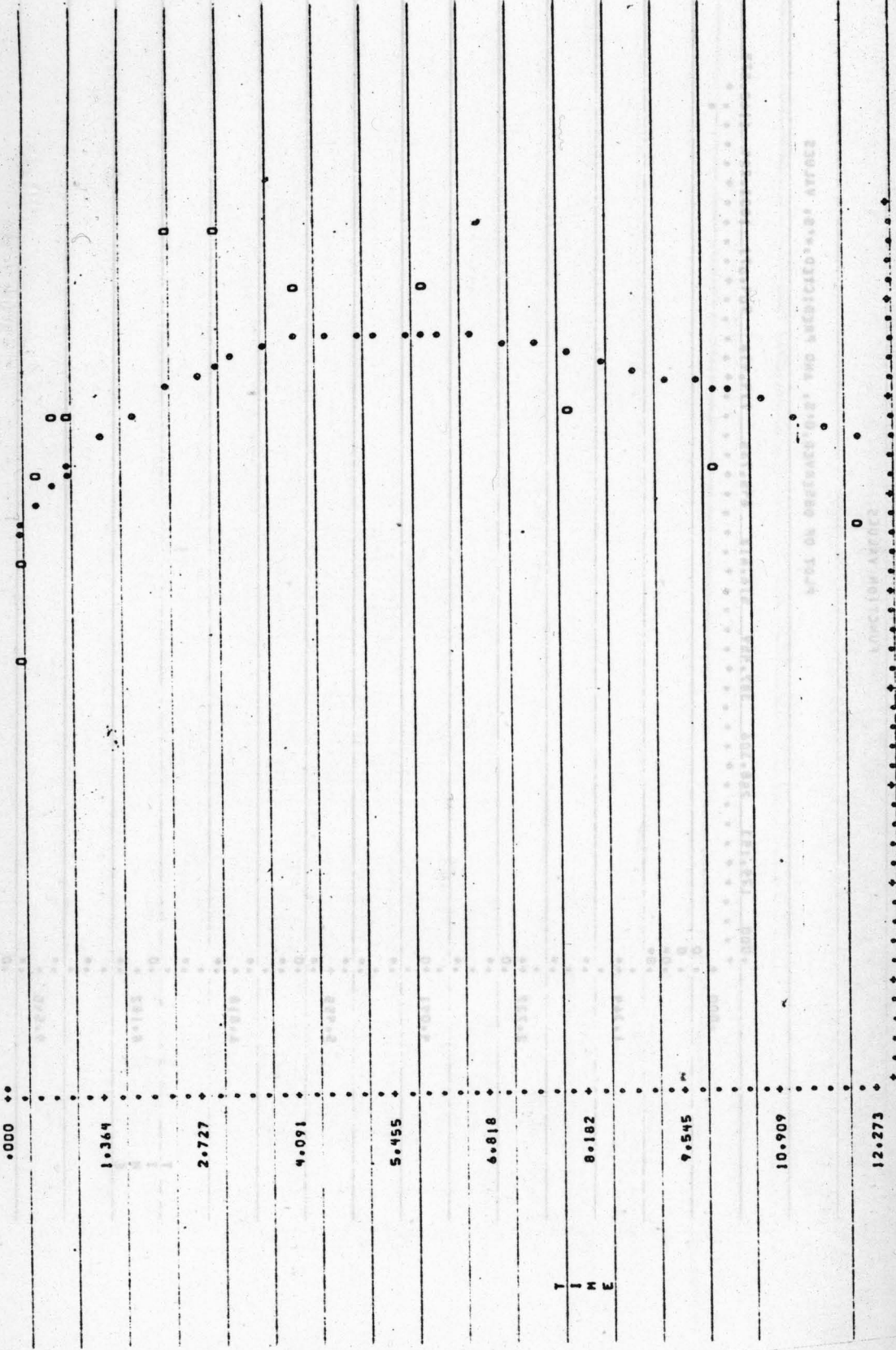


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1952-53 DEPARTMENT OF THE INTERIOR

PLOT OF OBSERVED, O'S, AND PREDICTED, P'S, VALUES

0.000 0.795 1.591 2.386 3.182 3.977 4.771 5.568 6.364 7.159



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APPL ON OBSERVED O'S AND PREDICTED P'S, VALUES

APPL ON OBSERVED O'S AND PREDICTED P'S, VALUES



SINGULAR VALUES AND VECTORS AT INITIAL POINT

SING. VALUES	8.1123259+00	3.9244547+00	8.8737120-01	2.7495076-01	6.6223234-02
1 KI	.0202595	-.0062094	.7552267	-.4793550	-.4464738
2 K-1	-.0170787	.0146188	-.5952069	-.7864457	-.1634846
3 K2	.4118137	-.9094622	-.0368103	-.0108664	-.0426010
4 KF	-.9108685	-.4112596	.0097965	.0111735	-.0310367
5 K3	...	...	...	...	...
6 DVI	-.0062094	.7552267	-.4793550	-.4464738	-.0000000
7 DVI	...	...	...	...	...

SETUP TIME = .170 SECONDS

1 1.0000000-01  
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SINGULAR VALUES AND VECTORS AT TERMINAL POINT

SING. VALUES	7.7353038+00	3.7183796+00	8.1893100+01	2.4159880+01	5.9728217+02
PAR. 1 K1	.0198493	-.0080514	.7691636	.4412928	-.4617241
2 K-1	-.0157299	.0159065	-.5674051	.8036389	-.1780882
3 K2	.4201985	-.9054582	-.0397039	-.0114706	-.0432506
4 KF	-.9070643	-.4195602	.0066415	-.0118357	-.0319262
5 K3	...	...	...	...	...
6 DV1	-.0080514	.7691636	.4412928	-.4617241	...
7 DV3	...	...	...	...	...

NORMALIZING ELEMENTS AND CORRELATION MATRIX

PARAMETER NO.	1	2	3	4	5
NORM. ELTS.	7.9986128+00	4.5204853+00	7.6889910+01	5.6095008+01	1.4618660+01
PAR. 1 K1	1.0000000	...	...	...	...
2 K-1	.7875051	1.0000000	...	...	...
3 K2	.8887915	.5850706	1.0000000	...	...
4 KF	.9026839	.5619391	.9508169	1.0000000	...
5 K3	...	...	...	...	...
6 DV1	.9313448	.5757798	.9436101	.9557852	1.0000000
7 DV3	...	...	...	...	...

PARAMETER NO. 7

PARAMETER NO.	1	2	3	4	5
NORM. ELTS.	...	...	...	...	...
PAR. 7 DV3	...	...	...	...	...

MADISON ACADEMIC COMPUTING CENTER - SUBROUTINE NREG  
METHUSOXIMIDE AND METABOLITE DATA 3-29-74 WEIGHTED BY 1/C  
07/30/75 TRANSMITTAL NO. B15365 CALL 1 PAGE 7

CONFIDENCE LIMITS ON LINEAR HYPOTHESIS

PAR.	LOWER LIMIT	FINAL VALUE	UPPER LIMIT
1 K1	5.1522264-01	3.8449433+00	7.1739638+00
2 K-1	1.9198263+00	3.8012512+00	5.6826760+00
3 K2	6.9140592-01	1.0114215+00	1.3314371+00
4 KF	5.3511557-01	7.6858284-01	1.0020501+00
F 5 K3	...	4.7000000-02	...
6 DV1	1.2842496+01	1.8926778+01	2.5011061+01
F 7 DV3	...	1.8900000+01	...

EXPLANATION

PAR.	LOWER TEST	SUM OF SQUARES	LINEAR EST. OF SUM OF SQUARES	UPPER TEST	SUM OF SQUARES	LINEAR EST. OF SUM OF SQUARES
1 K1	3.0126881+00	1.1088721+00	1.0796678+00	4.6771984+00	1.0387525+00	1.02796678+00
2 K-1	3.3308950+00	8.4464630-01	8.3968212-01	1.2716073+00	8.2846029-01	8.3968211-01
3 K2	9.3141763-01	9.3087831-01	9.1968346-01	1.0914254+00	9.0820169-01	9.1968346-01
4 KF	7.1021603-01	9.6126826-01	9.5550308-01	8.2694966-01	9.4384614-01	9.5550308-01
F 5 K3	4.7000000-02	...	...	4.7000000-02	...	...
6 DV1	1.7405708+01	1.0638779+00	1.0638658+00	2.0447849+01	1.0638607+00	1.0638658+00
F 7 DV3	1.8900000+01	...	...	1.8900000+01	...	...

TABLE 1007 REGRESSION

THIS IS THE STATE FULL OF THE DATA OF THE REGRESSION

MADISON ACADEMIC COMPUTING CENTER - SUBROUTINE NREG

FINAL FUNCTION VALUES AND RESIDUALS

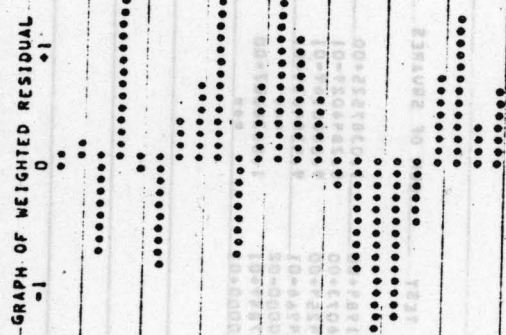
WEIGHTED ROOT MEAN SQUARE RESIDUAL = 2.0809987-01  
THIS IS THE SCALE UNIT IN THE GRAPH OF THE WEIGHTED RESIDUALS. NO. DEGREES OF FREEDOM = 18

EXPT. NO.	VAR. NO.	PREDICTION	OBSERVATION	RESIDUAL	WEIGHTED RES.
1	1	1.2665892+01	1.2600000+01	-6.5891623-02	-1.8520118-02
2	1	9.0445867+00	9.0999999+00	5.5432446-02	1.8378495-02
3	1	7.3194330+00	6.9000000+00	-4.1943300-01	-1.5971533-01
4	1	5.0485875+00	5.7000000+00	6.3141245-01	2.6413878-01
5	1	4.0582172+00	4.0000000+00	-5.8217200-01	-2.9108584-02
6	1	3.3242733+00	3.0000000+00	-3.2427328-01	-1.1830421-01
7	1	1.5186298+00	1.6000000+00	8.1370160-02	6.4328760-02
8	1	6.7418550-01	8.0000000-01	1.0581450-01	1.1830421-01
9	1	3.1732131-01	5.0000000-01	1.8267869-01	2.8884037-01
10	1	2.0182053+00	1.8000000+00	-2.1820536-01	-1.6270572-01
11	1	3.0558603+00	3.3000000+00	2.4413964-01	1.3438773-01
12	1	3.8238299+00	4.5000000+00	6.7617017-01	3.1859026-01
13	1	4.4320368+00	4.9000000+00	4.6796322-01	2.1136195-01
14	1	5.4320674+00	6.1000000+00	2.6793259-01	1.0850443-01
15	1	6.2972161+00	6.2000000+00	-9.7216129-02	-3.9007782-02
16	1	6.3431192+00	5.9000000+00	-4.4311923-01	-1.8216458-01
17	1	5.9901493+00	5.2000000+00	-7.9014927-01	-3.4622606-01
18	1	5.4978678+00	4.8000000+00	-5.9788781-01	-2.7004416-01
19	1	5.0140685+00	4.8000000+00	-2.1406653-01	-9.7630272-02
20	1	4.5661918+00	4.9000000+00	3.3380818-01	1.5076900-01
21	1	4.1569376+00	4.7000000+00	5.4306239-01	2.5063374-01
22	1	3.7840747+00	3.9000000+00	1.1592525-01	5.86654053-02
23	1	3.44445960+00	3.7000000+00	2.5540397-01	1.3271179-01

TIME SINCE END OF THE LAST ITERATION = .738 SECONDS TOTAL TIME = 2.107 SECONDS

R-SQUARED FOR FUNCTION1 = .9979  
COR FOR FUNCTION1 = .9951

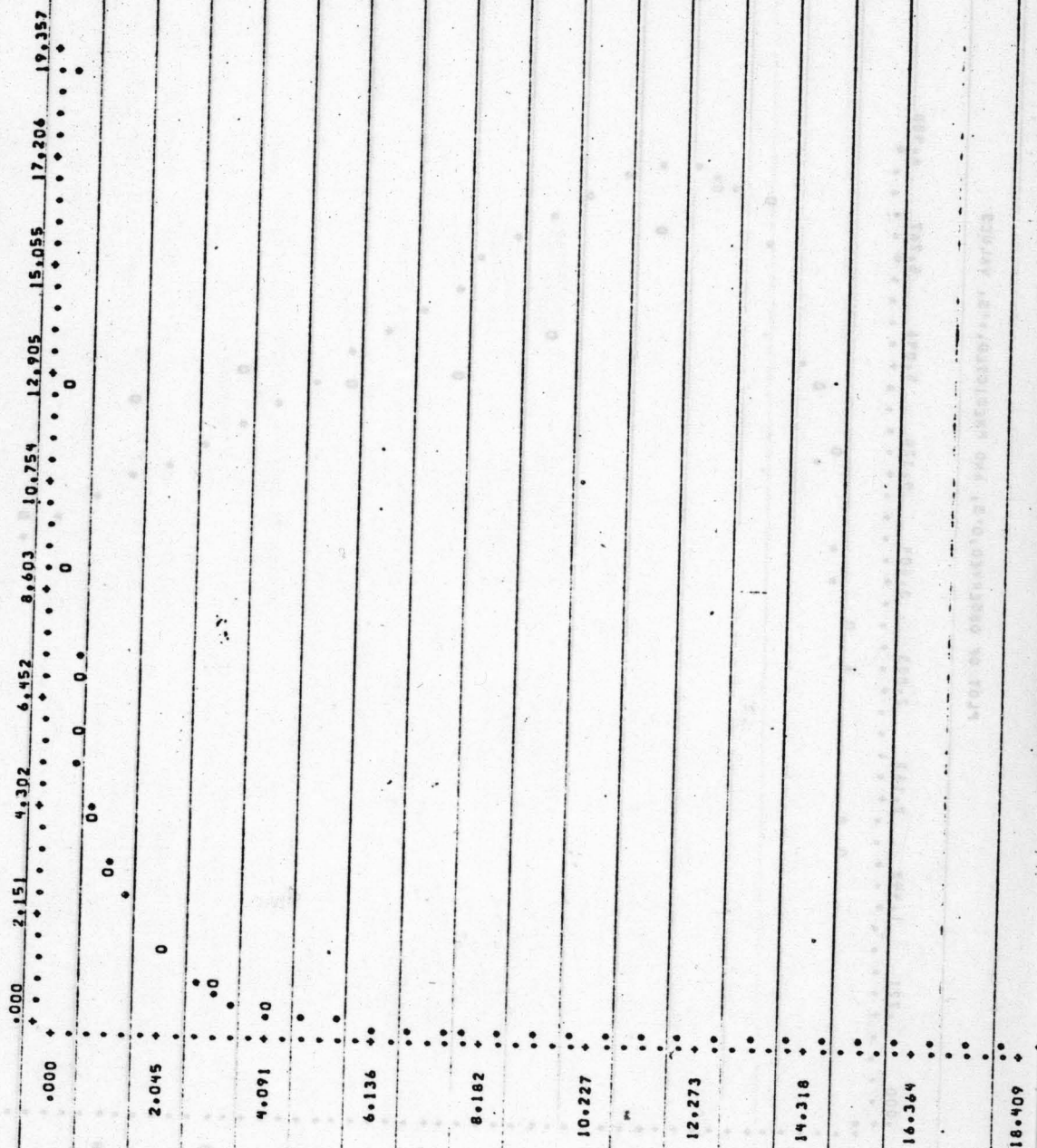
R-SQUARED FOR FUNCTION2 = .9919  
COR FOR FUNCTION2 = .9933  
OVER ALL R-SQUARED = .9950  
ALPHA = 8.64337  
BETA = .78282



HADISON ACADEMIC COMPUTING CENTER - SUBROUTINE MREG  
METHSUXIMIDE AND METABOLITE DATA 3-29-74 WEIGHTED BY 1/C

FUNCTION VALUES

PLOT OF OBSERVED, O'S, AND PREDICTED, P'S, VALUES

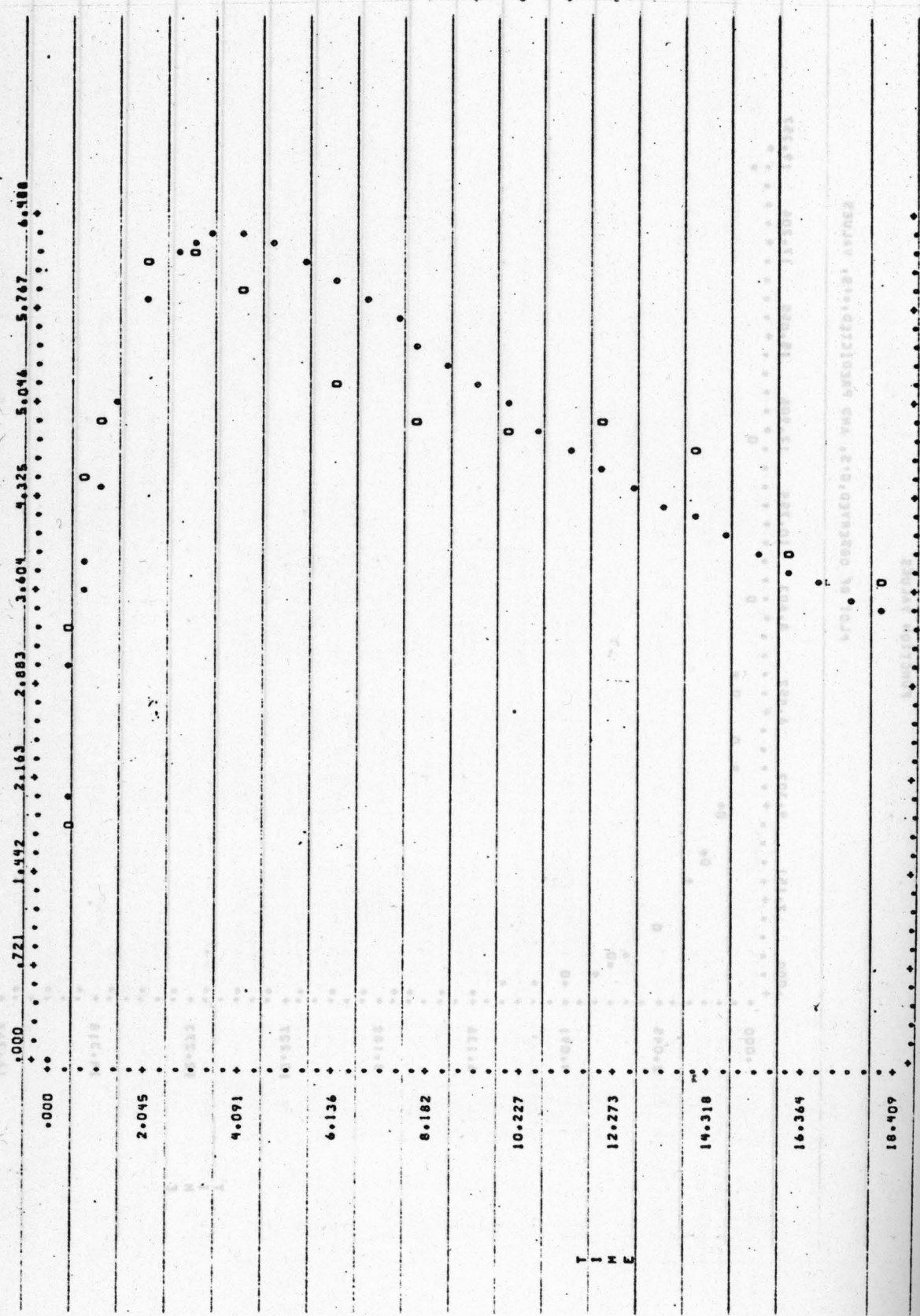


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FROM THE OBSERVATIONS AT THE UNIVERSITY OF CALIFORNIA

FUNCTION VALUES

PLOT OF OBSERVED, O'S, AND PREDICTED, P'S, VALUES



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FORM OBSERVED'S, P'S AND SPECIFIED'S, VALUES





THE ITERATIONS

ITERATION NO.	1	BASE POINT	TEST POINT
SUM OF SQUARES	1.1365013+00	1.0637526+00	0
LAMBDA	0.0000000	0.0000000	0
GAMMA	1.0000000+00	1.0000000+00	0
ANGLE IN DEGREES	0.0000	88.6763	0
MAX. PIVOT REDUCTION	1.0000000+00	5.2269500-02	0
PAR.			
1 K1	1.8600000+00	S	1.3704715+00
2 K-1	3.2800000+00		2.2616952+00
3 K2	5.5000000-01		5.5430590-01
4 KF	2.9000000-01		2.9586697-01
5 K3	F	4.7000000-02	F
6 DV1	1.1100000+01		1.1101266+01
7 DV3	F	1.8900000+01	F
CUMULATIVE NO. OF FUNCTION CALLS	2	ITERATION TIME	.188 SECONDS
			CUMULATIVE TIME
			.357 SECONDS

ITERATION NO.	10	BASE POINT	TEST POINT
SUM OF SQUARES	1.0152855+00	1.0152794+00	0
LAMBDA	0.0000000	0.0000000	0
GAMMA	1.0000000+00	1.0000000+00	0
ANGLE IN DEGREES	50.5741	52.5035	0
MAX. PIVOT REDUCTION	3.1164287-01	3.1303595-01	0
PAR.			
1 K1	S	7.1897584-01	S
2 K-1		1.1462594+00	7.1661857-01
3 K2		5.0498069-01	1.1419544+00
4 KF		2.7701470-01	5.0472535-01
5 K3	F	4.7000000-02	2.7691842-01
6 DV1		1.0438663+01	4.7000000-02
7 DV3	F	1.8900000+01	1.0435424+01
CUMULATIVE NO. OF FUNCTION CALLS	11	ITERATION TIME	.090 SECONDS
			CUMULATIVE TIME
			1.225 SECONDS

MADISON ACADEMIC COMPUTING CENTER - SUBROUTINE NREG  
METHSUXIMIDE AND METAROLITE DATA 6-5-74 WEIGHTED BY 1/C

THE LAST ITERATION

ITERATION NO.	15	BASE POINT	TEST POINT
SUM OF SQUARES	10	1.0152774+00	1.0152775+00
LAMBDA	10		
GAMMA	1.5392241+02	1.5108530+02	1.5108530+02
ANGLE IN DEGREES	33.2279	1.0000000+00	1.0000000+00
MAX. PIVOT REDUCTION	1.3877468+00	1.3806169+00	1.3806169+00

PAR.	1	K1	7.1464788-01
	2	K-1	1.1386636+00
	3	K2	5.0453240-01
	4	KF	2.7684771-01
	5	K3	4.7000000-02
	6	DV1	1.0433041+01
	7	DV3	1.8900000+01

CUMULATIVE NO. OF FUNCTION CALLS = 28

ITERATION TIME = 1.104 SECONDS

ITERATION TIME = 1.934 SECONDS

ITERATION TIME = 1.5000000-04

ITERATION TERMINATES:

MAX. RELATIVE CHANGE IN A PARAMETER .(L1, TOLL) = 1.5000000-04

FINAL PARAMETER VALUES

SUM OF SQUARES = 1.0152774\*00

AR.	FINAL VALUE
1 KI	7.1484788*01
2 K-1	1.1386636*00
3 K2	5.0453240*01
4 KF	2.7684771*01
5 K3	4.7000000*02
6 DVI	1.0433041*01
7 DV3	1.8900000*01

ITERATION SUMMARY

ITER NO.	SUM OF SQUARES	L	LAMBDA	GAMMA	ANGLE	NO. PIVOT FAILURES	NO. BOUNDS FAILURES	MAX. PIVOT REDUCTION	TIME IN SECONDS
0	1.1365013+00	0	0.0000000	1.0000000*00	.0000	0	0	1.0000000*00	.000
1	1.0637526+00	0	0.0000000	1.0000000*00	88.6463	0	0	5.2264950*02	.188
2	1.0407103+00	0	0.0000000	1.0000000*00	84.6198	0	0	2.0637511*01	.159
3	1.0259672+00	0	0.0000000	1.0000000*00	80.1231	0	0	2.3378379*01	.088
4	1.0191133+00	0	0.0000000	1.0000000*00	72.4771	0	0	2.6156288*01	.088
5	1.0165125+00	0	0.0000000	1.0000000*00	67.7928	0	0	2.8240300*01	.088
6	1.0156451+00	0	0.0000000	1.0000000*00	62.8085	0	0	2.9619780*01	.087
7	1.0153819+00	0	0.0000000	1.0000000*00	58.1573	0	0	3.049280*01	.089
8	1.0153076+00	0	0.0000000	1.0000000*00	55.1640	0	0	3.0916217*01	.089
9	1.0152855+00	0	0.0000000	1.0000000*00	50.5741	0	0	3.116428*01	.087
10	1.0152794+00	0	0.0000000	1.0000000*00	52.5035	0	0	3.1303595*01	.090
11	1.0152780+00	0	0.0000000	1.0000000*00	55.0506	0	0	3.1379437*01	.146
12	1.0152775+00	0	0.0000000	1.0000000*00	50.8098	0	0	3.1418685*01	.089
13	1.0152774+00	6	7.9434180*00	1.0000000*00	50.4615	0	0	7.6783449*01	.194
14	1.0152774+00	10	1.5392241*02	1.0000000*00	33.2379	0	0	1.3877468*00	.176
15	1.0152775+00	10	1.5108530*02	1.0000000*00	33.8369	0	0	1.3806167*00	.106

RELATIONSHIP AND METABOLITE DATA 6-5-74 WEIGHTED BY 1/C  
 MADISON ACADEMIC COMPUTING CENTER - SUBROUTINE NREG



CONFIDENCE LIMITS ON LINEAR HYPOTHESIS

PAR.	LOWER LIMIT	FINAL VALUE	UPPER LIMIT
1 K1	2.0365294-01	7.1484788-01	1.2260428+00
2 K-1	2.55970318-01	1.1386636+00	2.0214241+00
3 K2	4.2498590-01	5.0453240-01	5.8407889-01
4 KF	2.4023792-01	2.7684771-01	3.1315750-01
5 K3	..	4.7000000-02	..
6 DVI	9.3071828+00	1.0433041+01	1.1558900+01
7 DV3	..	1.8900000+01	..

EXPLORATION

PAR.	LOWER TEST	SUM. OF SQUARES	LINEAR EST. OF SUM OF SQUARES	UPPER TEST	SUM. OF SQUARES	LINEAR EST. OF SUM OF SQUARES
1 K1	5.8704915-01	1.0891182+00	1.0857894+00	8.4264661-01	1.0793176+00	1.0857894+00
2 K-1	9.1797353-01	1.0718083+00	1.0633190+00	1.3593538+00	1.0514349+00	1.0633190+00
3 K2	4.8464578-01	1.0485589+00	1.0472539+00	5.2441903-01	1.0459016+00	1.0472539+00
4 KF	2.6769526-01	1.0486782+00	1.0495286+00	2.8600015-01	1.0476126+00	1.0485286+00
5 K3	4.7000000-02	..	..	4.7000000-02	..	..
6 DVI	1.0151577+01	1.0497730+00	1.0497794+00	1.0714506+01	1.0497868+00	1.0497794+00
7 DV3	1.8900000+01	..	..	1.8900000+01	..	..

MADISON ACADEMIC COMPUTING CENTER - SUBROUTINE NREG  
 METHSUXIMIDE AND METABOLITE DATA 6-5-74 WEIGHTED BY 1/C

F I N A L F U N C T I O N V A L U E S A N D R E S I D U A L S

WEIGHTED ROOT MEAN SQUARE RESIDUAL = 2.0567747-01  
THIS IS THE SCALE UNIT IN THE GRAPH OF THE WEIGHTED RESIDUALS.  
NO. DEGREES OF FREEDOM = 24

EXPT. NO.	PREDICTION	OBSERVATION	RESIDUAL	WEIGHTED RES.	WEIGHTED RES.	GRAPH OF WEIGHTED RESIDUAL
1	9.9787569+00	1.070000+01	7.2124302-01	2.1994956-01	0	.....
2	9.2804216+00	8.599999+00	-6.8042171-01	-2.3174328-01	-1	.....
3	7.3662154+00	6.700000+00	-6.6621542-01	-2.5716260-01	0	.....
4	5.4804437+00	5.300000+00	-1.8044370-01	-7.8446331-02	0	.....
5	4.2823373+00	4.700000+00	4.1766268-01	1.9275936-01	0	.....
6	3.4886816+00	3.900000+00	4.1131839-01	2.0811247-01	0	.....
7	2.5317602+00	2.900000+00	3.6829981-01	2.1632716-01	0	.....
8	1.9680364+00	1.800000+00	-1.6803640-01	-1.2522702-01	0	.....
9	1.2821248+00	1.300000+00	1.7875180-02	1.5675218-02	0	.....
10	8.5712962-01	9.000000-01	4.2870179-02	4.5186876-02	0	.....
11	5.7541756-01	5.999999-01	2.4582438-02	3.1738964-02	0	.....
12	3.8655368-01	4.000000-01	1.3446320-02	2.1280498-02	0	.....
13	3.6691214-01	5.999999-01	2.3308786-01	3.0094522-01	0	.....
14	1.0177188+00	1.400000+00	3.8228121-01	3.2302197-01	0	.....
15	1.7424948+00	2.000000+00	2.5750524-01	1.8208370-01	0	.....
16	2.2834884+00	2.900000+00	6.1651158-01	3.6211856-01	0	.....
17	2.7046753+00	2.600000+00	-1.0467532-01	-6.4494329-02	0	.....
18	3.3246352+00	3.400000+00	7.5364292-02	4.0863851-02	0	.....
19	3.7616347+00	3.300000+00	-4.6163473-01	-2.5410884-01	0	.....
20	4.3145909+00	4.400000+00	8.5409045-02	4.0692727-02	0	.....
21	4.5967946+00	4.500000+00	-9.6794605-02	-4.4766554-01	0	.....
22	4.7076648+00	4.200000+00	-5.0766486-01	-2.4766554-01	0	.....
23	4.7077284+00	4.500000+00	-2.0772839-01	-9.7875126-02	0	.....
24	4.5215347+00	4.300000+00	-2.2153473-01	-1.0693498-01	0	.....
25	4.2224821+00	4.400000+00	1.7751783-01	8.457513-02	0	.....
26	3.8917753+00	4.400000+00	5.0824733-01	2.4214121-01	0	.....
27	2.2406474+00	2.400000+00	1.5932536-01	1.0288521-01	0	.....
28	1.2750088+00	1.300000+00	2.4991229-02	2.1915470-02	0	.....
29	7.2539110-01	5.000000-01	-2.62539110-01	-3.1875115-01	0	.....

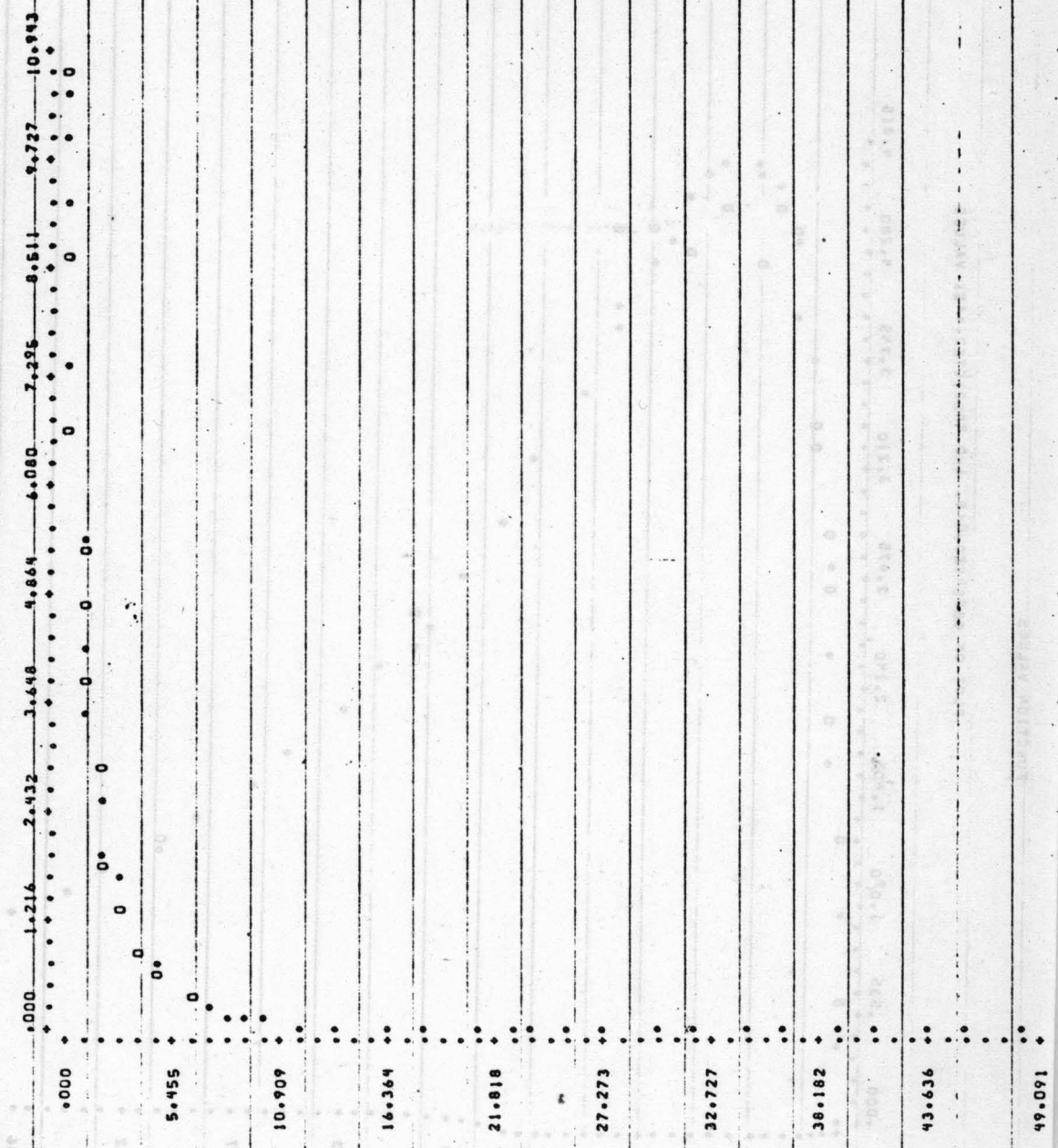
TIME SINCE END OF THE LAST ITERATION = .839 SECONDS TOTAL TIME = 2.775 SECONDS

R-SQUARED FOR FUNCTION1 = .9937  
COR FOR FUNCTION1 = .9931

R-SQUARED FOR FUNCTION2 = .9914  
COR FOR FUNCTION2 = .9928  
OVER ALL R-SQUARED = .9928  
ALPHA = 2.23719 BETA = .39770

FUNCTION VALUES

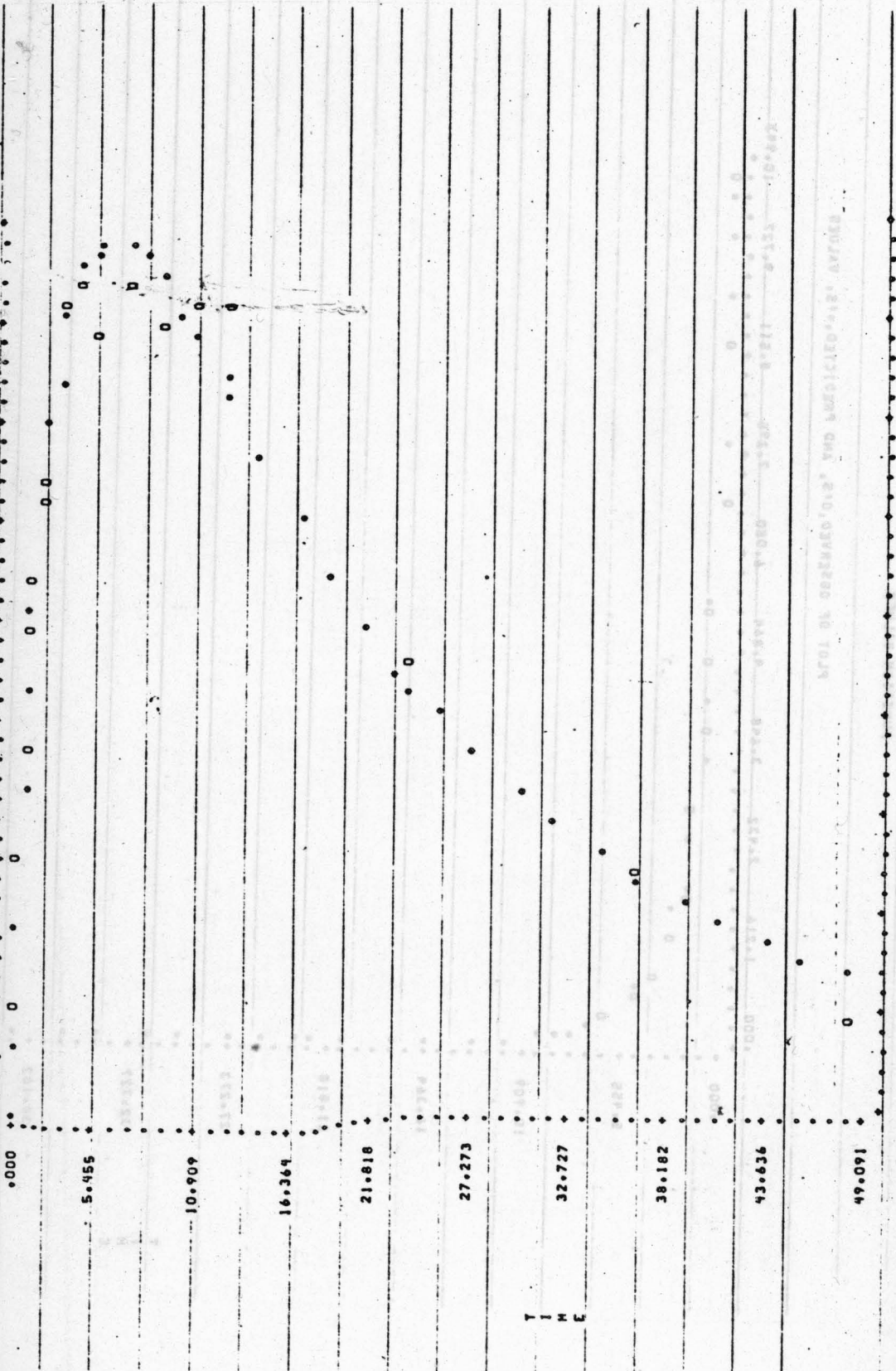
PLOT OF OBSERVED, O'S, AND PREDICTED, P'S, VALUES



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PLOT OF OBSERVED, O'S, AND PREDICTED, P'S, VALUES

.000 .535 1.070 1.605 2.140 2.675 3.210 3.745 4.280 4.815



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SCALING OF OBSERVED O'S AND PREDICTED P'S VALUES



## APPENDIX B

Derivation of Equations Describing Drug Blood Level vs  
Time Profiles Appropriate to Scheme III

The differential equations describing the amounts of drug in the central and tissue compartments and amount of metabolite in its central compartment as a function of time, appropriate to Scheme III, are:

$$\frac{dA_1}{dt} = k_{-1}A_2 - (k_1 + k_f + k_2)A_1 \quad \text{Eq. B1}$$

$$\frac{dA_2}{dt} = k_1A_1 - k_{-1}A_2 \quad \text{Eq. B2}$$

$$\frac{dM_b}{dt} = k_fA_1 - k_3M_b \quad \text{Eq. B3}$$

The application of the Laplace transform to linear differential equations is useful since it converts them into linear algebraic equations. The Laplace transform of a time function  $f(t)$  is  $F(s)$  as defined by equation B4,

$$F(s) = \int_0^{\infty} e^{-st} f(t) dt \quad \text{Eq. B4}$$

where  $s$  is a real number such that the integral converges for some finite value of  $s$  and all greater values. The Laplace transforms of functions common to pharmacokinetics do not have to be calculated with equation B4 each time but can be found in Laplace transform tables (21).

The Laplace transforms of equations B1-B3 are

$$(s + k_1 + k_f + k_2)\bar{A}_1 + (-k_{-1})\bar{A}_2 = D \quad \text{Eq. B5}$$

$$(-k_1)\bar{A}_1 + (s + k_{-1})\bar{A}_2 = 0 \quad \text{Eq. B6}$$

$$(-k_f)\bar{A}_1 + (s + k_3)\bar{M}_b = 0 \quad \text{Eq. B7}$$

where the bar represents the Laplace transform of that quantity and D is the dose of methsuximide.

This set of linear algebraic equations (B5-B7) can be solved for  $\bar{A}_1$ ,  $\bar{A}_2$  and  $\bar{M}_b$  by matrix algebra utilizing Cramer's rule. Hence, the matrix of coefficients is

$$\Delta = \begin{vmatrix} (s + k_1 + k_f + k_2) & -k_{-1} & 0 \\ -k_1 & (s + k_{-1}) & 0 \\ -k_f & 0 & (s + k_3) \end{vmatrix} \quad \text{Eq. B8}$$

which upon expansion becomes

$$\Delta = (s + k_3) \left\{ s^2 + (k_1 + k_f + k_2 + k_{-1})s + k_{-1}k_f + k_2k_{-1} \right\} \quad \text{Eq. B9}$$

If  $\alpha$  and  $\beta$  are defined as

$$\frac{\alpha}{\beta} = \frac{1}{2} \left\{ (k_1 + k_f + k_2 + k_{-1}) \pm \right. \quad \text{Eq. B10}$$

$$\left. \sqrt{(k_1 + k_f + k_2 + k_{-1})^2 - 4(k_{-1}k_f + k_2k_{-1})} \right\}$$

then equation B9 reduces to B11.

$$\Delta = (s + k_3)(s + \alpha)(s + \beta) \quad \text{Eq. B11}$$

The  $\bar{A}_1$  matrix is given by equation B12

$$\Delta \bar{A}_1 = \begin{vmatrix} D & -k_{-1} & 0 \\ 0 & (s + k_{-1}) & 0 \\ 0 & 0 & (s + k_3) \end{vmatrix} \quad \text{Eq. B12}$$

which upon expansion becomes

$$\Delta \bar{A}_1 = D(s + k_{-1})(s + k_3) \quad \text{Eq. B13}$$

The value of the Laplace transformed  $A_1$  is then

$$\bar{A}_1 = \frac{\Delta \bar{A}_1}{\Delta} \quad \text{Eq. B14}$$

$$\text{or } \bar{A}_1 = \frac{Ds + Dk_{-1}}{(s + \alpha)(s + \beta)} \quad \text{Eq. B15}$$

The anti-Laplace of equation B15, found in Tables (21), is

$$A_1 = \frac{(Dk_{-1} - D\alpha)e^{-\alpha t} - (Dk_{-1} - D\beta)e^{-\beta t}}{\beta - \alpha} \quad \text{Eq. B16}$$

which rearranges to equation B17, the desired equation, which describes the amount of methsuximide in the central compartment as a function of time, i.e. text equation 19.

$$A_1 = \frac{D}{(\alpha - \beta)} \left\{ (k_{-1} - \beta)e^{-\beta t} - (k_{-1} - \alpha)e^{-\alpha t} \right\} \quad \text{Eq. B17}$$

Utilizing a similar development (equations B18-B20) yields equation B21 which describes the amount of methsuximide in the tissue compartment as a function of time, i.e. text equation 20

$$\Delta \bar{A}_2 = \begin{vmatrix} (s + k_1 + k_f + k_2) & D & 0 \\ -k_1 & 0 & 0 \\ -k_f & 0 & (s + k_3) \end{vmatrix} \quad \text{Eq. B18}$$

$$\Delta \bar{A}_2 = Dk_1 (s + k_3) \quad \text{Eq. B19}$$

$$\bar{A}_2 = \frac{\Delta \bar{A}_2}{\Delta} = \frac{Dk_1}{(s + \alpha)(s + \beta)} \quad \text{Eq. B20}$$

$$A_2 = \frac{Dk_1}{(\alpha - \beta)} (e^{-\beta t} - e^{-\alpha t}) \quad \text{Eq. B21}$$

Similarly, the development (equations B22-B24) for 2-methyl-2-phenylsuccinimide results in equation B25, i.e. text equation 21

$$\Delta \bar{M}_b = \begin{vmatrix} (s + k_1 + k_f + k_2) & -k_{-1} & D \\ -k_1 & (s + k_{-1}) & 0 \\ -k_f & 0 & 0 \end{vmatrix} \quad \text{Eq. B22}$$

$$\Delta \bar{M}_b = Dk_f (s + k_{-1}) \quad \text{Eq. B23}$$

$$\bar{M}_b = \frac{\Delta \bar{M}_b}{\Delta} = \frac{Dk_f (s + k_{-1})}{(s + k_3) (s + \alpha) (s + \beta)} \quad \text{Eq. B24}$$

$$M_b = Dk_f \left\{ \left[ \frac{\alpha - k_{-1}}{(k_3 - \alpha) (\alpha - \beta)} \right] e^{-\alpha t} + \right. \quad \text{Eq. B25}$$

$$\left. \left[ \frac{\beta - k_{-1}}{(\beta - k_3) (\alpha - \beta)} \right] e^{-\beta t} + \left[ \frac{k_3 - k_{-1}}{(\beta - k_3) (k_3 - \alpha)} \right] e^{-k_3 t} \right\}$$

It is to be noted that the units of D, the dose of methsuximide, should be in  $\mu\text{moles}$  since biotransformation of methsuximide to 2-methyl-2-phenylsuccinimide is on a mole for mole basis not  $\mu\text{g}$  for  $\mu\text{g}$ . However, in these experiments concentrations were measured as  $\mu\text{g ml}^{-1}$ , dose was in mg and distribution volumes were in liters. Therefore, the dose in mg was prefixed by the ratio of molecular weights of metabolite to methsuximide (189/203) so that  $C_{M_b}$  units of  $\mu\text{g ml}^{-1}$  could be used. When the computer uses equation B26 to calculate  $C_{M_b}$  with D in mg and V in l the unit will be  $\mu\text{g ml}^{-1}$ .

$$C_{M_b} = \frac{189Dk_f}{203V_3} \left\{ \left[ \frac{\alpha - k_{-1}}{(k_3 - \alpha) (\alpha - \beta)} \right] e^{-\alpha t} + \right. \quad \text{Eq. B26}$$

$$\left. \left[ \frac{\beta - k_{-1}}{(\beta - k_3) (\alpha - \beta)} \right] e^{-\beta t} + \left[ \frac{k_3 - k_{-1}}{(\beta - k_3) (k_3 - \alpha)} \right] e^{-k_3 t} \right\}$$

## APPENDIX C

Area Under the Metabolite Curve After  
Intravenous Methsuximide Dosing

Equation C1 gives the concentration of the metabolite in the body after an intravenous methsuximide dose as a function of time.

$$C_{M_b} = \frac{189}{203} \frac{Dk_f}{V_3} \left\{ \left[ \frac{\alpha - k_{-1}}{(k_3 - \alpha)(\alpha - \beta)} \right] e^{-\alpha t} + \left[ \frac{\beta - k_{-1}}{(\beta - k_3)(\alpha - \beta)} \right] e^{-\beta t} + \left[ \frac{k_3 - k_{-1}}{(\alpha - k_3)(k_3 - \alpha)} \right] e^{-k_3 t} \right\} \quad \text{Eq. C1}$$

The area under this curve is given by equation C2 which becomes equation C3 after integration.

$$\begin{aligned} \text{Area } C_{M_b} &= \frac{189k_f D}{203V_3} \left\{ \int_0^{\infty} \frac{(\alpha - k_{-1})}{(k_3 - \alpha)(\alpha - \beta)} e^{-\alpha t} dt + \int_0^{\infty} \frac{(\beta - k_{-1})}{(\beta - k_3)(\alpha - \beta)} e^{-\beta t} dt + \int_0^{\infty} \frac{(k_3 - k_{-1})}{(\beta - k_3)(k_3 - \alpha)} e^{-k_3 t} dt \right\} \quad \text{Eq. C2} \end{aligned}$$

$$\begin{aligned} \text{Area } C_{M_b} &= \frac{189k_f D}{203V_3} \left\{ \frac{1}{\alpha} \frac{(\alpha - k_{-1})}{(k_3 - \alpha)(\alpha - \beta)} + \frac{1}{\beta} \frac{(\beta - k_{-1})}{(\alpha - \beta)(\beta - k_3)} + \frac{1}{k_3} \frac{(k_3 - k_{-1})}{(\beta - k_3)(k_3 - \alpha)} \right\} \quad \text{Eq. C3} \end{aligned}$$

When the quantities within the bracket in equation C3 are put over the common denominator and simplified equation C4 results (text equation 26),

$$\text{Area } C_{M_b} = \frac{189}{203} \frac{D}{V_3} \frac{k_f^{k-1}}{k_3 \alpha \beta} \quad \text{Eq. C4}$$

where D is the dose of methsuximide in mg,  $V_3$  is the distribution volume of the metabolite in l and all rate constants have units of  $\text{hour}^{-1}$ . The units of area  $C_{M_b}$  is then  $\mu\text{g hr ml}^{-1}$ .

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